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(54) Recombinant polypeptides and their uses, including assay for AIDS virus.

(57) Novel recombinant HTLV-III envelope proteins denoted R10, PB1, 590, and KH1, are useful in the diagnosis, prophylaxis or therapy of AIDS. Protein R10 is a 95 kD fusion protein; protein PB1 is a 26 kD fusion protein; protein 590 is an 86 kD fusion protein; and protein KH1 is a 70 kD fusion protein. These proteins are considered to be especially useful to prepare vaccines for the HTLV-III virus.

EP 0 255 190 A2

RECOMBINANT POLYPEPTIDES AND THEIR USES,
INCLUDING ASSAY FOR AIDS VIRUS

This invention relates to recombinant polypeptides and their uses, including assay for AIDS virus. The virus which causes AIDS (acquired immune deficiency syndrome) has been identified as human T-cell lymphotropic virus type III (HTLV-III), lymphadenopathy-associated virus (LAV) or AIDS-associated retrovirus (ARV); see Popovic et al, Science 224 (1984) 497-500. The virus has been designated HIV (human immunodeficiency virus) by the International Committee on Taxonomy of Virus.

The virus displays tropism for the OKT4⁺ lymphocyte subset; see Klatzmann et al, Science 225 (1984) 59-63.

Antibodies against HTLV-III proteins in the sera of most AIDS and AIDS-related complex (ARC) patients, and in asymptomatic people infected with the virus (Sarngadharan et al, Science 224 (1984) 506-508) have made possible the development of immunologically based tests that detect antibodies to these antigens. These tests are used to limit the spread of HTLV-III through blood transfusion by identifying blood samples of people infected with the virus. Diagnostic tests currently available commercially use the proteins of inactivated virus as antigens.

In addition to allowing new approaches for diagnosis, recombinant DNA holds great promise for the development of vaccines against both bacteria and viruses; see Wilson, Bio/Technology 2 (1984) 29-39.

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-2-

The most widely employed organisms to express recombinant vaccines have been E. coli, S. cerevisiae and cultured mammalian cells. For example, subunit vaccines against foot and mouth disease (Kleid, D.G.,
5 Yansura, D., Small, B., Dowbenko, D., Moore, D.M., Brubman, M.J., McKercher, P.D., Morgan, D.O., Robertson, B.H. and Bachrach, H.L. [1981] Science 214:1125-1129) and malaria (Young, J.F., Hockmeyer, W.T., Gross, M.,
10 Ripley Ballou, W., Wirtz, R.A., Trosper, J.H., Beaudoin, R.L., Hollingdale, M.R., Miller, L.M., Diggs, C.L. and Rosenberg, M. [1985] Science 228:958-962) have been synthesized in E. coli. Other examples are hepatitis B surface antigen produced in yeast (McAleer, W.J., Buynak, E.B., Maigetter, R.Z., Wampler, D.E.,
15 Miller, W.J. and Hilleman, M.R. [1984] Nature 307: 178-180) and a herpes vaccine produced in mammalian cells (Berman, P.W., Gregory, T., Chase, D. and Lasky, L.A. [1984] Science 227:1490-1492).

20 There is a real need at this time to develop a vaccine for AIDS. No such vaccine is known to exist.

Brief Summary of the Invention

25 The subject invention concerns novel recombinant HTLV-III proteins and the uses thereof. More specifically, the subject invention concerns four novel recombinant HTLV-III envelope proteins which can be used in the diagnosis, prophylaxis or therapy of AIDS.
30 Further, the recombinant HTLV-III envelope protein fragments of the invention can be used to stimulate a lymphocyte proliferative response in HTLV-III infected humans. This then would stimulate the immune system to respond to HTLV-III in such individuals and,
35 therefore, the envelope protein fragments can provide protection and be of therapeutic value.

These novel proteins are encoded on bacterial plasmids which can be used to transform suitable hosts, for example, E. coli, using standard procedures.

In the accompanying drawings:

5 Figures 1A and 1B are sequential flow charts of the construction, from plasmid pBG1, of plasmid pREV2.2 which is used to construct vectors encoding novel proteins;

Figure 2 is a diagram of plasmid pREV2.2 and also of the multiple cloning site; and

10 Figure 3 is a schematic representation of the HTLV-III envelope gene and also of novel recombinant proteins obtained therefrom.

Expression vector plasmid pREV2.2 was constructed from plasmid pBG1 by the route shown in Figure 1 of the
15 drawings. In the product, the hatched region represents TrpA trase. terminator and the dotted region represents multiple cloning sites for enzymes NruI, MluI, EcoRV, ClaI, BamHI, SalI, HindIII, SmaI, StsI, EcoRI.

Plasmid pR10 contains approximately 1275 base pairs
20 of DNA encoding the HTLV-III env gene from essentially the KpnI site to the BglII. This plasmid in a suitable bacterial host, e.g. E. coli, can be used to produce the novel recombinant HTLV-III 95 kD fusion protein denoted R10. The amino-acid sequence of fusion protein R10 is
25 shown in Table 8; the DNA sequence encoding this protein is shown in Table 8A.

Plasmid pPB1 contains approximately 540 base pairs of DNA encoding essentially the HTLV-III env gene from the PvuII site to the BglII site. This plasmid in a
30 suitable host, e.g. E. coli, can be used to produce the novel recombinant HTLV-III 26 kD fusion protein denoted PB1. The amino-acid sequence of fusion protein PB1 is shown in Table 9; the DNA sequence encoding this protein is shown in Table 9A.

Plasmid p590 contains approximately 1055 base pairs of DNA encoding essentially the HTLV-III env gene from the PvuII site to the HindIII site. This plasmid in a suitable host, e.g., E. coli, can be used to produce the novel recombinant HTLV-III 86 kD protein denoted 590. The amino acid sequence of fusion protein 590 is shown in Table 10; the DNA sequence encoding this protein is shown in Table 10A.

Plasmid pKH1 contains approximately 1830 base pairs of DNA encoding essentially the HTLV-III env gene from the KpnI site to the HindIII site. This plasmid in a suitable host, e.g., E. coli, can be used to produce the novel recombinant HTLV-III 70 kD protein denoted KH1. The amino acid sequence of fusion protein KH1 is shown in Table 11; the DNA sequence encoding this protein is shown in Table 11A.

Plasmid pBG1 is deposited in the E. coli host MS371 with the Northern Regional Research Laboratory (NRRL), U.S. Department of Agriculture, Peoria, Illinois, USA. It is in the permanent collection of this repository. E. coli MS371(pBG1), NRRL B-15904, was deposited on Nov. 1, 1984. E. coli MS371, NRRL B-15129, is now available to the public. E. coli SG20251, NRRL B-15918, was deposited on Dec. 12, 1984.

Other relevant NRRL deposits, their deposit dates and numbers, are as follows:

Culture	Accession No.	Date of Deposit
<u>E. coli</u> JM103(pREV2.2)	NRRL B-18091	July 30, 1986
<u>E. coli</u> SG20251(pR10)	NRRL B-18093	July 30, 1986
<u>E. coli</u> SG20251(pPB1)	NRRL B-18092	July 30, 1986
<u>E. coli</u> SG20251(p590)	NRRL B-18094	July 30, 1986
<u>E. coli</u> CAG629(pKH1)	NRRL B-18095	July 30, 1986

The novel HTLV-III proteins of the subject invention can be expressed in Saccharomyces cerevisiae using plasmids containing the inducible galactose promoter from this organism (Broach, J.R., Li, Y.,
5 Wu, L.C. and Jayaram, M. in Experimental Manipulation of Gene Expression [1983] p. 83, ed. M. Inouye, Academic Press). These plasmids are called YEp51 and YEp52 (Broach, J.R. et al. [1983]) and contain the
10 E. coli origin of replication, the gene for β -lactamase, the yeast LEU2 gene, the 2 μ m origin of replication and the 2 μ m circle REP3 locus. Recombinant gene expression is driven by the yeast GAL10 gene promoter.

Yeast promoters such as galactose and alcohol dehydrogenase (Bennetzen, J.L. and Hall, B.D. [1982]
15 J. Biol. Chem. 257:3018; Ammerer, G. in Methods in Enzymology [1983] Vol. 101, p. 192), phosphoglycerate kinase (Derynck, R., Hitzeman, R.A., Gray, P.W.,
Goeddel, D.V., in Experimental Manipulation of Gene Expression [1983] p. 247, ed. M. Inouye, Academic
20 Press), triose phosphate isomerase (Alber, T. and Kawasaki, G. [1982] J. Molec. and Applied Genet. 1:419), or enolase (Innes, M.A. et al. [1985] Science 226:21) can be used.

The genes disclosed herein can be expressed in simian cells. When the genes encoding these proteins are cloned into one of the plasmids as described in Okayama and Berg (Okayama, H. and Berg, P. [1983] Molec. and Cell. Biol. 3:280) and references therein, or COS cells transformed with these plasmids, synthesis of HTLV-III proteins can be detected immunologically.

Other mammalian cell gene expression/protein production systems can be used. Examples of other such systems are the vaccinia virus expression system (Moss, B. [1985] Immunology Today 6:243; Chakrabarti, S., Brechling, K., Moss, B. [1985] Molec. and Cell. Biol. 5:3403) and the vectors derived from murine retroviruses (Mulligan, R.C. in Experimental Manipulation of Gene Expression [1983] p. 155, ed. M. Inouye, Academic Press).

The HTLV-III proteins of the subject invention can be chemically synthesized by solid phase peptide synthetic techniques such as BOC and FMOC (Merrifield, R.B. [1963] J. Amer. Chem. Soc. 85:2149; Chang, C. and Meienhofer, J. [1978] Int. J. Peptide Protein Res. 11:246).

As is well known in the art, the amino acid sequence of a protein is determined by the nucleotide sequence of the DNA. Because of the redundancy of the genetic code, i.e., more than one coding nucleotide triplet (codon) can be used for most of the amino acids used to make proteins, different nucleotide sequences can code for a particular amino acid. Thus, the genetic code can be depicted as follows:

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-7-

	Phenylalanine (Phe)	TTK	Histidine (His)	CAK
	Leucine (Leu)	XTY	Glutamine (Gln)	CAJ
	Isoleucine (Ile)	ATM	Asparagine (Asn)	AAK
	Methionine (Met)	ATG	Lysine (Lys)	AAJ
5	Valine (Val)	GTL	Aspartic acid (Asp)	GAK
	Serine (Ser)	QRS	Glutamic acid (Glu)	CAJ
	Proline (Pro)	CCL	Cysteine (Cys)	TGK
	Threonine (Thr)	ACL	Tryptophan (Trp)	TGG
	Alanine (Ala)	GCL	Arginine (Arg)	WGZ
10	Tyrosine (Tyr)	TAK	Glycine (Gly)	GGL
	Termination Signal	TAJ		
	Termination Signal	TGA		

Key: Each 3-letter deoxynucleotide triplet corresponds to a trinucleotide of mRNA, having a 5'-end on the left and a 3'-end on the right. All DNA sequences given herein are those of the strand whose sequence corresponds to the mRNA sequence, with thymine substituted for uracil. The letters stand for the purine or pyrimidine bases forming the deoxynucleotide sequence.

A = adenine

G = guanine

C = cytosine

T = thymine

25 X = T or C if Y is A or G

X = C if Y is C or T

Y = A, G, C or T if X is C

Y = A or G if X is T

W = C or A if Z is A or G

30 W = C if Z is C or T

Z = A, G, C or T if W is C

Z = A or G if W is A

QR = TC if S is A, G, C or T; alternatively QR =
AG if S is T or C

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J = A or G
K = T or C
L = A, T, C or G
M = A, C or T

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The above shows that the novel amino acid sequences of the HTLV-III proteins of the subject invention can be prepared by nucleotide sequences other than those disclosed herein. Functionally equivalent nucleotide sequences encoding the novel amino acid sequences of these HTLV-III proteins, or fragments thereof having HTLV-III antigenic or immunogenic or therapeutic activity, can be prepared by known synthetic procedures. Accordingly, the subject invention includes such functionally equivalent nucleotide sequences.

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Thus the scope of the subject invention includes not only the specific nucleotide sequences depicted herein, but also all equivalent nucleotide sequences coding for molecules with substantially the same HTLV-III antigenic or immunogenic or therapeutic activity.

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Further, the scope of the subject invention is intended to cover not only the specific amino acid sequences disclosed, but also similar sequences coding for proteins or protein fragments having comparable ability to induce the formation of and/or bind to specific HTLV-III antibodies.

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The term "equivalent" is being used in its ordinary patent usage here as denoting a nucleotide sequence which performs substantially as the nucleotide sequence identified herein to produce molecules with substantially the same HTLV-III antigenic or immunogenic or therapeutic activity in essentially the same kind of hosts. Within this definition are subfragments which have HTLV-III antigenic or immunogenic or therapeutic activity.

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-9-

As disclosed above, it is well within the skill of those in the genetic engineering art to use the nucleotide sequences encoding HTLV-III antigenic or immunogenic or therapeutic activity of the subject invention to produce HTLV-III proteins via microbial processes. Fusing the sequences into an expression vector and transforming or transfecting into hosts, either eukaryotic (yeast or mammalian cells) or prokaryotic (bacterial cells), are standard procedures used in producing other well-known proteins, e.g., insulin, interferons, human growth hormone, IL-1, IL-2, and the like. Similar procedures, or obvious modifications thereof, can be employed to prepare HTLV-III proteins by microbial means or tissue-culture technology in accord with the subject invention.

The nucleotide sequences disclosed herein can be prepared by a "gene machine" by procedures well known in the art. This is possible because of the disclosure of the nucleotide sequence.

The restriction enzymes disclosed can be purchased from Bethesda Research Laboratories, Gaithersburg, MD, or New England Biolabs, Beverly, MA. The enzymes are used according to the instructions provided by the supplier.

The various methods employed in the preparation of the plasmids and transformation of host organisms are well known in the art. These procedures are all described in Maniatis, T., Fritsch, E.F., and Sambrook, J. (1982) Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York. Thus, it is within the skill of those in the genetic engineering

-10-

art to extract DNA from microbial cells, perform restriction enzyme digestions, electrophorese DNA fragments, tail and anneal plasmid and insert DNA, ligate DNA, transform cells, e.g., E. coli cells, prepare plasmid DNA, electrophorese proteins, and sequence DNA.

Immunochemical assays employing the HTLV-III proteins of the invention can take a variety of forms. The preferred type is a solid phase immunometric assay. In assays of this type, an HTLV-III protein is immobilized on a solid phase to form an antigen-immunoabsorbent. The immunoabsorbent is incubated with the sample to be tested. After an appropriate incubation period, the immunoabsorbent is separated from the sample and labeled anti-(human IgG) antibody is used to detect human anti-HTLV-III antibody bound to the immunoabsorbent. The amount of label associated with the immunoabsorbent can be compared to positive and negative controls to assess the presence or absence of anti-HTLV-III antibody.

The immunoabsorbent can be prepared by adsorbing or coupling a purified HTLV-III protein to a solid phase. Various solid phases can be used, such as beads formed of glass, polystyrene, polypropylene, dextran or other material. Other suitable solid phases include tubes or plates formed from or coated with these materials.

The HTLV-III proteins can be either covalently or non-covalently bound to the solid phase by techniques such as covalent bonding via an amide or ester linkage or adsorption. After the HTLV-III protein is affixed to the solid phase, the solid phase can be post-coated with an animal protein, e.g., 3% fish gelatin. This provides a blocking protein which reduces nonspecific

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-11-

adsorption of protein to the immunoabsorbent surface.

5 The immunoabsorbent is then incubated with the sample to be tested for anti-HTLV-III antibody. In blood screening, blood plasma or serum is used. The plasma or serum is diluted with normal animal plasma or serum. The diluent plasma or serum is derived from the same animal species that is the source of the anti-(human IgG) antibody. The preferred anti-(human IgG) antibody is goat anti-(human IgG) antibody. Thus, in the preferred format, the diluent would be goat serum or plasma.

10 The conditions of incubation, e.g., pH and temperature, and the duration of incubation are not crucial. These parameters can be optimized by routine experimentation. Generally, the incubation will be run for 1-2 hr at about 45°C in a buffer of pH 7-8.

15 After incubation, the immunoabsorbent and the sample are separated. Separation can be accomplished by any conventional separation technique such as sedimentation or centrifugation. The immunoabsorbent then may be washed free of sample to eliminate any interfering substances.

20 The immunoabsorbent is incubated with the labeled anti-(human IgG) antibody (tracer) to detect human antibody bound thereto. Generally the immunoabsorbent is incubated with a solution of the labeled anti-(human IgG) antibody which contains a small amount (about 1%) of the serum or plasma of the animal species which serves as the source of the anti-(human IgG) antibody. Anti-(human IgG) antibody can be obtained from any animal source. However, goat anti-(human IgG) antibody is preferred. The anti-(human IgG) antibody can be an

-12-

antibody against the Fc fragment of human IgG, for example, goat anti-(human IgG) Fc antibody.

5 The anti-(human IgG) antibody or anti-(human IgG)Fc can be labeled with a radioactive material such as ¹²⁵iodine; labeled with an optical label, such as a fluorescent material; or labeled with an enzyme such as horseradish peroxidase. The anti-human antibody can also be biotinylated and labeled avidin used to detect its binding to the immunoadsorbent.

10 After incubation with the labeled antibody, the immunoadsorbent is separated from the solution and the label associated with the immunoadsorbent is evaluated. Depending upon the choice of label, the evaluation can be done in a variety of ways. The label
15 may be detected by a gamma counter if the label is a radioactive gamma emitter, or by a fluorimeter, if the label is a fluorescent material. In the case of an enzyme, label detection may be done colorimetrically employing a substrate for the enzyme.

20 The amount of label associated with the immunoadsorbent is compared with positive and negative controls in order to determine the presence of anti-HTLV-III antibody. The controls are generally run concomitantly with the sample to be tested. A positive
25 control is a serum containing antibody against HTLV-III; a negative control is a serum from healthy individuals which does not contain antibody against HTLV-III.

30 For convenience and standardization, reagents for the performance of the immunometric assay can be assembled in assay kits. A kit for screening blood, for example, can include:

- (a) an immunoadsorbent, e.g., a polystyrene bead coated with an HTLV-III protein;

-13-

- 5 (b) a diluent for the serum or plasma sample,
e.g., normal goat serum or plasma;
- (c) an anti-(human IgG) antibody, e.g., goat
anti-(human IgG) antibody in buffered, aqueous
solution containing about 1% goat serum or
plasma;
- 10 (d) a positive control, e.g., serum containing
antibody against at least one of the novel
HTLV-III proteins; and
- (e) a negative control, e.g., pooled sera from
healthy individuals which does not contain
antibody against at least one of the novel
HTLV-III proteins.

15 If the label is an enzyme, an additional element
of the kit can be the substrate for the enzyme.

Another type of assay for anti-HTLV-III antibody
is an antigen sandwich assay. In this assay, a labeled
HTLV-III protein is used in place of anti-(human IgG)
antibody to detect anti-HTLV-III antibody bound to the
immunoabsorbent. The assay is based in principle on the
bivalency of antibody molecules. One binding site of
the antibody binds the antigen affixed to the solid
phase; the second is available for binding the labeled
antigen. The assay procedure is essentially the same
as described for the immunometric assay except that
after incubation with the sample, the immunoabsorbent
is incubated with a solution of labeled HTLV-III protein.
HTLV-III proteins can be labeled with radioisotope,
an enzyme, etc. for this type of assay.

30 In a third format, the bacterial protein, protein A,
which binds the Fc segment of an IgG molecule without
interfering with the antigen-antibody interaction can
be used as the labeled tracer to detect anti-HTLV-
antibody adsorbed to the immunoabsorbent. Protein A

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-14-

can be readily labeled with a radioisotope, enzyme or other detectable species.

5 Immunochemical assays employing an HTLV-III protein have several advantages over those employing a whole (or disrupted) virus. Assays based upon an HTLV-III protein will alleviate the concern over growing large quantities of infectious virus and the inherent variability associated with cell culturing and virus production. Further, the assay will help mitigate the
10 real or perceived fear of contracting AIDS by technicians in hospitals, clinics and blood banks who perform the test.

 Vaccines of the HTLV-III proteins, disclosed herein, and variants thereof having antigenic properties,
15 can be prepared by procedures well known in the art. For example, such vaccines can be prepared as injectables, e.g., liquid solutions or suspensions. Solid forms for solution in, or suspension in, a liquid prior to injection also can be prepared. Optionally, the
20 preparation also can be emulsified. The active antigenic ingredient or ingredients can be mixed with excipients which are pharmaceutically acceptable and compatible with the active ingredient. Examples of suitable excipients are water, saline, dextrose, glycerol,
25 ethanol, or the like, and combinations thereof. In addition, if desired, the vaccine can contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, or adjuvants which enhance the effectiveness of the vaccine. The
30 vaccines are conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for other modes of administration include suppositories and, in some cases, oral formulations.

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-15-

For suppositories, traditional binders and carriers include, for example, polyalkylene glycols or triglycerides. Suppositories can be formed from mixtures containing the active ingredient in the range of about 0.5% to about 10%, preferably about 1 to about 2%.

Oral formulations can include such normally employed excipients as, for example, pharmaceutical grades of manitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate and the like. These compositions can take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain from about 10% to about 95% of active ingredient, preferably from about 25% to about 70%.

The proteins can be formulated into the vaccine as neutral or salt forms. Pharmaceutically acceptable salts include the acid addition salts (formed with the free amino groups of the peptide) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups also can be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, procaine, and the like.

The vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective and immunogenic. The quantity to be administered depends on the subject to be treated, capacity of the subject's immune system to synthesize antibodies, and the degree of protection desired. Precise amounts of active ingredient required

to be administered depend on the judgment of the practitioner and are peculiar to each individual. However, suitable dosage ranges are of the order of about several hundred micrograms active ingredient per individual. Suitable regimes for initial administration and booster shots are also variable, but are typified by an initial administration followed in one or two week intervals by a subsequent injection or other administration.

HTLV-III is known to undergo amino acid sequence variation, particularly in the envelope gene (Starcich, B.R. [1986] Cell 45:637-648; Hahn, B.H. et al. [1986] Science 232:1548-1553). Over 100 variants have been analyzed by molecular cloning and restriction enzyme recognition analysis, and several of these have been analyzed by nucleotide sequencing. Some of these are the HTLV-III isolates known as RF (Popovic, M. et al. [1984] Science 224:497-500), WMJ-1 (Hahn, B.H. et al. [1986] Science 232:1548-1553), LAV (Wain-Hobson, S. et al. [1985] Cell 40:9-17), and ARV-2 (Sanchez-Pescador, R. et al. [1985] Science 227:484-492). Although the subject invention describes the sequence from one HTLV-III isolate, the appropriate envelope regions of any HTLV-III isolate can be produced using the procedures described herein for preparing R10, PB1, 590, and KH1. The HTLV-III proteins from different viral isolates can be used in vaccine preparations, as disclosed above, to protect against infections by different HTLV-III isolates. Further, a vaccine preparation can be

-17-

made using more than one recombinant antigenic protein from more than one HTLV-III isolate to provide immunity and thus give better protection against AIDS.

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Following are examples which illustrate the process of the invention, including the best mode. These examples should not be construed as limiting. All solvent mixture proportions are by volume unless otherwise noted.

10

Example 1--Construction of plasmid pREV2.2

The pREV2.2 plasmid expression vector was constructed from plasmid pBG1. Plasmid pBG1 can be isolated from its E. coli host by well known procedures, e.g., using cleared lysate-isopycnic density gradient procedures, and the like. Like pBG1, pREV2.2 expresses inserted genes behind the E. coli promoter. The differences between pBG1 and pREV2.2 are the following:

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1. pREV2.2 lacks a functional replication of plasmid (rop) protein.
2. pREV2.2 has the trpA transcription terminator inserted into the AatII site. This sequence insures transcription termination of over-

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-18-

3. pREV2.2 has genes to provide resistance to ampicillin and chloramphenicol, whereas pBG1 provides resistance only to ampicillin.
4. pREV2.2 contains a sequence encoding sites for several restriction endonucleases.

The following procedures, shown in Figure 1 of the drawings, were used to make each of the four changes listed above:

- 1a. 5 μ g of plasmid pBG1 was restricted with NdeI which gives two fragments of approximately 2160 and 3440 base pairs.
- 1b. 0.1 μ g of DNA from the digestion mixture, after inactivation of the NdeI, was treated with T4 DNA ligase under conditions that favor intramolecular ligation (200 μ l reaction volume using standard T4 ligase reaction conditions [New England Biolabs, Beverly, MA]). Intramolecular ligation of the 3440 base pair fragment gave an ampicillin resistant plasmid. The ligation mixture was transformed into the recipient strain E. coli JM103 (available from New England Biolabs) and ampicillin resistant clones were selected by standard procedures.
- 1c. The product plasmid, pBG1 Δ N, where the 2160 base pair NdeI fragment is deleted from pBG1, was selected by preparing plasmid from ampicillin resistant clones and determining

-19-

the restriction digestion patterns with NdeI and Sall (product fragments approximately 1790 and 1650). This deletion inactivates the rop gene that controls plasmid replication.

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2a. 5 μ g of pBG1 Δ N was then digested with EcoRI and BclI and the larger fragment, approximately 2455 base pairs, was isolated.

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2b. A synthetic double stranded fragment was prepared by the procedure of Itakura et al. (Itakura, K., Rossi, J.J. and Wallace, R.B. [1984] Ann. Rev. Biochem. 53:323-356, and references therein) with the structure shown in Table 1. This fragment has BclI and EcoRI sticky ends and contains recognition sequences for several restriction endonucleases.

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2c. 0.1 μ g of the 2455 base pair EcoRI-BclI fragment and 0.01 μ g of the synthetic fragment were joined with T4 DNA ligase and competent cells of strain JM103 were transformed. Cells harboring the recombinant plasmid, where the synthetic fragment was inserted into pBG1 Δ N between the BclI and EcoRI sites, were selected by digestion of the plasmid with HpaI and EcoRI. The diagnostic fragment sizes are approximately 2355 and 200 base pairs. This plasmid is called pREV1.

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2d. 5 μ g of pREV1 were digested with AatII, which cleaves uniquely.

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2e. The double stranded fragment shown in Table 2 was synthesized. This fragment has AatII sticky ends and contains the trpA transcription termination sequence.

2f. 0.1 μ g of AatII digested pREV1 was ligated with 0.01 μ g of the synthetic fragment in a volume

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-20-

of 20 μ l using T4 DNA ligase.

2g. Cells of strain JM103, made competent, were transformed and ampicillin resistant clones selected.

5 2h. Using a KpnI, EcoRI double restriction digest of plasmid isolated from selected colonies, a cell containing the correct construction was isolated. The sizes of the KpnI, EcoRI generated fragments are approximately 2475 and 80 base pairs. This plasmid is called pREV1TT and contains the trpA transcription terminator.

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3a. 5 μ g of pREV1TT, prepared as disclosed above (by standard methods) was cleaved with NdeI and XmnI and the approximately 850 base pair fragment was isolated.

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3b. 5 μ g of plasmid pBR325 (BRL, Gaithersburg, MD), which contains the genes conferring resistance to chloramphenicol as well as to ampicillin and tetracycline, was cleaved with BclI and the ends blunted with Klenow polymerase and deoxynucleotides. After inactivating the enzyme, the mixture was treated with NdeI and the approximately 3185 base pair fragment was isolated. This fragment contains the genes for chloramphenicol and ampicillin resistance and the origin of replication.

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3c. 0.1 μ g of the NdeI-XmnI fragment from pREV1TT and the NdeI-BclI fragment from pBR325 were ligated in 20 μ l with T4 DNA ligase and the mixture used to transform competent cells of strain JM103. Cells resistant to both ampicillin and chloramphenicol were selected.

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-21-

- 3d. Using an EcoRI and NdeI double digest of plasmid from selected clones, a plasmid was selected giving fragment sizes of approximately 2480, 1145, and 410 base pairs. This is called plasmid pREVI^{TT}/chl and has genes for resistance to both ampicillin and chloramphenicol.
- 4a. A double stranded fragment shown in Table 3 was synthesized. This fragment, with a blunt end and an SstI sticky end, contains recognition sequences for several restriction enzyme sites.
- 4b. 5 µg of pREVI^{TT}/chl was cleaved with NruI (which cleaves about 20 nucleotides from the BclI site) and SstI (which cleaves within the multiple cloning site). The larger fragment, approximately 3990 base pairs, was isolated from an agarose gel.
- 4c. 0.1 µg of the NruI-SstI fragment from pREVI^{TT}/chl and 0.01 µg of the synthetic fragment were treated with T4 DNA ligase in a volume of 20 µl.
- 4d. This mixture was transformed into strain JM103 and ampicillin resistant clones were selected.
- 4e. Plasmid was purified from several clones and screened by digestion with MluI or ClaI. Recombinant clones with the new multiple cloning site will give one fragment when digested with either of these enzymes, because each cleaves the plasmid once.
- 4f. The sequence of the multiple cloning site was verified. This was done by restricting the

-22-

plasmid with HpaI and PvuII and isolating the 1395 base pair fragment, cloning it into the SmaI site of mpl8 and sequencing it by dideoxynucleotide sequencing using standard methods.

5 4g. This plasmid, called pREV2.2 is diagrammed in Figure 2 of the drawings.

Example 2--Construction of and expression from pR10

10 Plasmid pR10, which contains approximately 1275 base pairs of DNA encoding the HTLV-III env gene from essentially the KpnI site to the BglIII site, and from which is synthesized an approximately 95 kD fusion protein containing this portion of the gp120 envelope
15 protein, can be constructed as follows:

1. Synthesizing the DNA with the sequence shown in Table 4. This DNA fragment can be synthesized by standard methods (Itakura, et al., supra, and references therein) and encodes
20 a portion of gp120. It has a blunt end on the 5' end and an end which will ligate with a BamHI overhang on the 3' end.
2. Restricting 5 µg of plasmid pBG1 with BclI, filling in the overhanging ends with Klenow polymerase and deoxyribonucleotide triphosphates (dNTPs), restricting this fragment
25 with BamHI and isolating the large fragment, approximately 8.9 kb, from an agarose gel.
3. Ligating 0.1 µg of the fragment in Table 4 with
30 0.1 µg of the pBG1 fragment in a volume of 20 µl using T4 DNA ligase, transforming the ligation mixture into competent cell strain SG20251 (Gottesman, S., Halbern, E. and Trisler, P. [1981] Journal of Bacteriology 148:265-273),
35 and selecting ampicillin resistant transformants.

-23-

4. Selecting, using the AhaIII restriction pattern of purified plasmid, cells harboring the recombinant plasmid with the synthesized fragment in the orientation whereby the fragment blunt end ligated to the pBG1 fragment filled-in BclI end and the BamHI overhanging ends ligated together. AhaIII digestion of the proper plasmid gives fragment lengths of approximately 5300, 3170, 690, 640, 330, and 20 base pairs.
5. When the strain harboring this recombinant plasmid is grown in 2% medium (2% yeast extract, bactotryptone, casamino acids (Difco, Detroit, MI), 0.2% potassium monobasic, 0.2% potassium dibasic, and 0.2% sodium dibasic) containing 50 µg/ml ampicillin and the total complement of cellular proteins electrophoresed on an SDS-polyacrylamide gel, a prominent protein of approximately 95 kD can be visualized by either coomassie blue staining or by western blot analysis using as probe selected sera from AIDS, ARC, or HTLV-III infected individuals.

Example 3--Purification of recombinant protein containing HTLV-III envelope sequences from plasmid pR10

1. Growth of cells:

Cells were grown in a 10 liter volume in a Chemap fermentor (Chemapec, Woodbury, NY) in 2% medium. Fermentation temperature was 37°C, the pH was 6.8, and air was provided at 1 vvm. Plasmid selection was provided by 50 µg/ml ampicillin. Typical cell yield (wet weight) is 30 g/l.

2. Cell lysis:

50 g, wet cell weight, of E. coli containing the recombinant HTLV-III envelope fusion protein were

resuspended in a final volume of 100 ml in 50 mM Tris-Cl pH 8.0, 5 mM potassium ethylenediaminetetraacetic acid (KEDTA), 5 mM dithiothreitol (DTT), 15 mM β -mercaptoethanol, 0.5% Triton X-100, and 5 mM phenylmethylsulfonyl fluoride (PMSF). 300 mg lysozyme was added and the suspension incubated for 30 min at room temperature.

This material was lysed using a BEAD-BEATERTM (Biospec Products, Bartlesville, OK) containing an equal volume of 0.1-0.15 μ m glass beads. The lysis was done for 6 min at room temperature in 1 min intervals. The liquid was separated from the beads and centrifuged for 2.5 hr at 20,000 xg. The supernatant was removed and the pellet dissolved in 100 ml 8 M urea, 20 mM Tris-Cl pH 8.0, 5 mM DTT, 15 mM β -mercaptoethanol, 5 mM PMSF, and 1 mM KEDTA. The pellet was solubilized using a polytron homogenizer (Beckman, Berkeley, CA) and centrifuged at 20,000 xg for 2 hr.

3. Diethylaminoethyl (DEAE) chromatography:

Supernatant was loaded onto a 550 ml column (5 cm x 28 cm) packed with DEAE Fast Flow Sepharose (Pharmacia, Piscataway, NJ) equilibrated in 8 M urea, 20 mM Tris-Cl pH 8.0, 15 mM β -mercaptoethanol, and 1 mM KEDTA at room temperature. The column was washed with 1.5 liters equilibration buffer, and the protein eluted with a 5.0 liter linear gradient from 0-0.8 M NaCl in equilibration buffer. The HTLV-III protein eluted at 0.2 M NaCl and was assayed using SDS-polyacrylamide electrophoresis and following the prominent protein at approximately 95 kD.

The fractions containing the HTLV-III protein were pooled and the protein concentrated to 10 ml using a stressed cell positive pressure concentrator

-25-

(Amicon, Danvers, MA) fitted with a 10,000 MW cut-off membrane (YM-10, Amicon). The concentrate was loaded onto a 500 ml column (2.5 cm x 100 cm) packed with superfine sephacryl S-300 (Pharmacia) equilibrated in 8 M urea, 20 mM Tris-Cl, pH 8.0, 15 mM β -mercaptoethanol, and 1 mM KEDTA. The column was eluted with equilibration buffer at room temperature. A flow rate of 0.5 ml/min was maintained. The HTLV-III protein eluted at approximately 40% of the column volume.

Example 4--Construction of and expression from plasmid pPB1

Plasmid pPB1, which contains approximately 540 base pairs of DNA encoding essentially the HTLV-III env gene from the PvuII site to the BglIII site, and from which is synthesized an approximately 26 kD fusion protein containing this portion of the gp120 envelope protein can be constructed as follows:

1. Synthesizing the DNA with the sequence shown in Table 5: This DNA fragment can be synthesized by standard methods and encodes a portion of gp120. It has a blunt end on the 5' end and an end which will ligate with a BamHI overhang on the 3' end.
2. Restricting 5 μ g plasmid pREV2.2 with EcoRV and BamHI and isolating the large fragment, approximately 4 kD, from an agarose gel.
3. Ligating 0.1 μ g of the fragment in Table 5 with 0.1 μ g of the pREV2.2 fragment in a volume of 20 μ l using T4 DNA ligase, transforming the ligation mixture into competent cell strain SG20251, and selecting ampicillin resistant transformants.
4. Using the AhaIII restriction pattern of purified plasmid, selecting cells harboring the recombinant

-26-

plasmid with the synthesized fragment in the orientation whereby the fragment blunt end ligated to the REV2.2 EcoRV end and the BamHI overhanging ends ligated together. AhaIII digestion of the proper plasmid gives fragment lengths of approximately 1210, 1020, 750, 690, 500, 340, and 20 base pairs. When the strain harboring this recombinant plasmid is grown in 2% medium containing 50 µg/ml ampicillin and the total complement of cellular proteins electrophoresed on an SDS-polyacrylamide gel, a protein of approximately 26 kD can be visualized by either coomassie blue staining or by western blot analysis using as probe selected sera from AIDS, ARC, or HTLV-III infected individuals.

Example 5--Purification of recombinant protein containing HTLV-III envelope sequences from plasmid pPB1

1. Growth of cells:

Cells were grown in a 10 liter volume in a Chemap fermentor in 2% medium. Fermentation temperature was 37°C, the pH was 6.8, and air was provided at 1 vvm. Plasmid selection was provided by 50 µg/ml ampicillin and 20 µg/ml chloramphenicol. Typical cell yield (wet weight) was 30 g/l.

2. Cell Lysis:

50 g, wet cell weight, of E. coli containing the recombinant HTLV-III envelope fusion protein were resuspended in a final volume of 100 ml in 50 mM Tris-Cl pH 8.0, 5 mM KEDTA, 5 mM DTT, 15 mM β-mercaptoethanol, 0.5% Triton X-100, and 5 mM PMSF. 300 mg lysozyme was added and the suspension incubated for 30 min at room temperature.

This material was lysed using a BEAD-BEATERTM (Biospec Products, Bartlesville, OK) containing an

-27-

equal volume of 0.1-0.15 μ m glass beads. The lysis was done for 6 min at room temperature in 1 min intervals. The liquid was separated from the beads and centrifuged for 2.5 hr at 20,000 xg. The supernatant was removed and the pellet was resuspended in 100 ml 6 M guanidine-hydrochloride, 20 mM Tris-Cl pH 8.0, 5 mM DTT, 15 mM β -mercaptoethanol, 5 mM PMSF, and 1 mM KEDTA. The pellet was solubilized using a polytron homogenizer and centrifuged at 20,000 xg for 2 hr.

The supernate (90 ml) was dialysed against 4 liters of 8 M urea, 20 mM Tris-Cl, pH 8.0, 1 mM EDTA, and 15 mM β -mercaptoethanol. Dialysis was done each time for 8 hr or longer with three changes of buffer. Spectraphor dialysis tubing (S/P, McGraw Park, IL) with a 3.5 kD MW cut-off was used.

3. CM chromatography

The dialysate was loaded onto a 550 ml column (5 cm x 28 cm) packed with CM Fast Flow Sepharose (Pharmacia) equilibrated in 8 M urea, 10 mM potassium phosphate pH 7.0, 15 mM β -mercaptoethanol, and 1 mM KEDTA at room temperature. The column was washed with 2 liters equilibration buffer, and the protein eluted with a 5.0 liter linear gradient from 0-0.4 M NaCl. The HTLV-III protein (26 kD) eluted at approximately 0.2 M NaCl as assayed by SDS-polyacrylamide gel electrophoresis.

Example 6--Construction of and expression from plasmid p590

Plasmid p590, which contains approximately 1055 base pairs of DNA encoding essentially the HTLV-III

-28-

env gene from the PvuII site to the HindIII site, and from which is synthesized an approximately 86 kD fusion protein containing this portion of the gp160 envelope protein can be constructed as follows:

- 5 1. Synthesizing the DNA with the sequence shown in Table 6: This DNA fragment can be synthesized by standard methods and encodes a portion of gp160. It has a blunt end on the 5' end and an end which will ligate with a HindIII overhang on the 3' end.
- 10 2. Restricting 5 µg plasmid pREV2.2 with EcoRV and HindIII and isolating the large fragment, approximately 4 kD, from an agarose gel.
- 15 3. Ligating 0.1 µg of the fragment in Table 6 with 0.1 µg of the pREV2.2 fragment in a volume of 20 ml using T4 DNA ligase, transforming the ligation mixture into competent cell strain SG20251, and selecting ampicillin resistant transformants.
- 2- 4. Using the AhaIII restriction pattern of purified plasmid, selecting cells harboring the recombinant plasmid with the synthesized fragment in the orientation whereby the fragment blunt end ligated to the pREV2.2 EcoRV end and the HindIII overhanging ends ligated together. AhaIII digestion of the proper plasmid gives fragment lengths of approximately 1740, 1020, 750, 690, 500, 340, and 20.
- 25 5. 5 µg of plasmid, purified from this strain, is restricted with NdeI and SmaI. The approximately 1425 base pair fragment is isolated from an agarose gel. The 1505 base pair fragment is fused to the DNA encoding the segment of gp160.
- 30 6. 5 µg of pBG101 is restricted with BamHI, the overhanging ends filled in with Klenow polymerase and dNTPs, and then restricted with NdeI. The approximately 6.5 kD fragment is isolated from an agarose gel.
- 35

-29-

7. Ligating 0.1 μ g of the NdeI-SmaI fragment with 0.1 μ g of the pBG1 fragment using T4 DNA ligase, transforming the ligation mixture into competent cell strain SG20251, and selecting ampicillin resistant transformants.
8. Using the AhaIII restriction pattern of purified plasmid, selecting cells harboring the recombinant plasmid with the synthesized fragment in the orientation whereby the fragment blunt SmaI end ligated to the BamHI/filled-in end and the NdeI overhanging ends ligated together. AhaIII digestion of the proper plasmid gives fragment lengths of approximately 5900, 1020, 690, 430, and 20 base pairs.
9. When the strain harboring this recombinant plasmid is grown in 2% medium containing 50 μ g/ml ampicillin and the total complement of cellular proteins electrophoresed on an SDS-polyacrylamide gel, a protein of approximately 86 kD can be visualized by either coomassie blue staining or by western blot analysis using as probe selected sera from AIDS, ARC, or HTLV-III infected individuals.

Example 7--Purification of recombinant protein containing HTLV-III envelope sequences from plasmid p590

1. Growth of cells:

Cells were grown in a 10 liter volume in a Chemap fermentor in 2% medium. Fermentation temperature was 37°C, the pH was 6.8, and air was provided at 1 vvm. Plasmid selection was provided by 50 μ g/ml ampicillin. Typical cell yield (wet weight) is 30 g/l.

2. Cell Lysis:

50 g, wet cell weight, of E. coli containing the recombinant HTLV-III envelope fusion protein were

-30-

resuspended in a final volume of 100 ml in 50 mM Tris-Cl pH 8.0, 5 mM KEDTA, 5 mM DTT, 15 mM β -mercaptoethanol, 0.5% Triton X-100, and 5 mM PMSF. 300 mg lysozyme was added and the suspension incubated for 30 min at room temperature.

This material was lysed using a Bead-BeaterTM containing 0.1-0.15 mm glass beads. The lysis was done for 6 min at room temperature in 1 min intervals. The liquid was separated from the beads and centrifuged for 2.5 hr at 20,000 xg. The supernatant was removed and the pellet was resuspended in 100 ml 6 M guanidine-hydrochloride, 20 mM Tris-Cl pH 8.0, 5 mM DTT, 15 mM β -mercaptoethanol, 5 mM PMSF, and 1 mM KEDTA. The pellet was solubilized using a polytron homogenizer and centrifuged at 20,000 xg for 2 hr.

The supernate (90 ml) was dialysed against 4 liters of 8 M urea, 20 mM Tris-Cl pH 8.0, 1 mM EDTA, and 15 mM β -mercaptoethanol. Dialysis was done each time for 8 hr or longer with three changes of buffer. Spectrophor dialysis tubing with a 3.5 kD MW cut-off was used.

3. Diethylaminoethyl (DEAE) chromatography

Dialysate was loaded onto a 550 ml column (5 cm x 28 cm) packed with DEAE Fast Flow Sepharose (Pharmacia) equilibrated in 8 M urea, 20 mM Tris-Cl pH 8.0, 15 mM β -mercaptoethanol, and 1 mM KEDTA at room temperature. The column was washed with 1.5 liters equilibration buffer, and the protein eluted with a 5.0 liter linear gradient from 0-0.8 M NaCl in equilibration buffer. The HTLV-III protein eluted at 0.4 M NaCl and was assayed using SDS-polyacrylamide electrophoresis and following the prominent protein at approximately 86 kD.

-31-

The fractions containing the HTLV-III protein were pooled and the protein concentrated to 10 ml using a stressed cell positive pressure concentrator (Amicon) fitted with a 10,000 MW cut-off membrane (YM-10, Amicon). The concentrate was loaded onto a 500 ml column (2.5 cm x 100 cm) packed with superfine sephacryl S-300 (Pharmacia) equilibrated in 8 M urea, 20 mM Tris-Cl, pH 8.0, 15 mM β -mercaptoethanol, and 1 mM KEDTA. The column was eluted with equilibration buffer at room temperature. A flow rate of 0.5 ml/min was maintained. The HTLV-III protein eluted at approximately 40% of the column volume.

Example 8--Construction of and expression from plasmid pKH1

Plasmid pKH1, which contains approximately 1830 base pairs of DNA encoding essentially the HTLV-III env gene from the KpnI site to the HindIII site, and from which is synthesized an approximately 70 kD fusion protein containing this portion of the gpl60 envelope protein, can be constructed as follows:

1. Synthesizing the DNA with the sequence shown in Table 7: This DNA fragment can be synthesized by standard methods and encodes a portion of gpl60. It has a blunt end on the 5' end and an end which will ligate with a HindIII overhang on the 3' end.
2. Restricting 5 μ g plasmid pREV2.2 with MluI, treating the DNA with Klenow polymerase to blunt the ends, treating with HindIII and isolating the large fragment, approximately 5 kD, from an agarose gel.
3. Ligating 0.1 μ g of the fragment in Table 7 with 0.1 μ g of the pREV 2.2 fragment in a

-32-

volume of 20 μ l using T4 DNA ligase, transforming the ligation mixture into competent cell strain CAG629, and selecting ampicillin resistant transformants.

- 5 4. Using the AhaIII restriction pattern of purified plasmid, selecting cells harboring the recombinant plasmid with the synthesized fragment in the orientation whereby the fragment blunt end ligated to the REV2.2 MluI
- 10 end and the HindIII overhanging ends ligated together. AhaIII digestion of the proper plasmid gives fragment lengths of approximately 1730, 1020, 750, 690, 640, 600, 340, and 20
- 15 base pairs. When the strain harboring this recombinant plasmid is grown in 2% medium containing 50 μ g/ml ampicillin and the total complement of cellular proteins electrophoresed on an SDS-polyacrylamide gel, a protein of approximately 70 kD can be visualized by either
- 20 Coomassie blue staining or by Western Blot analysis using as probe selected sera from AIDS, ARC, or HTLV-III infected individuals.

25 Example 9--Purification of recombinant protein containing HTLV-III envelope sequences from plasmid pKH1

1. Growth of cells:

Cells were grown in a 10 liter volume in a Chemap fermenter in 2% medium. Fermentation temperature was 32°C, the pH was 6.8, and air was provided at 1 vvm. Plasmid selection was provided by 50 μ g/ml ampicillin. Typical cell yield (wet weight) is 30 g/l.

-33-

2. Cell lysis:

50 g, wet cell weight, of E. coli containing the recombinant HTLV-III envelope fusion protein were resuspended in a final volume of 100 ml in 50 mM Tris-Cl pH 8.0, 5 mM KEDTA, 5 mM dithiothreitol (DTT), 15 mM β -mercaptoethanol, 0.5% Triton X-100 and 5 mM PMSF. 300 mg lysozyme was added and the suspension incubated for 30 min at room temperature.

This material was lysed using a BEAD-BEATERTM (Biospec Products) containing an equal volume of 0.1-0.15 μ m glass beads. The lysis was done for 6 min at room temperature in 1 min intervals. The liquid was separated from the beads and centrifuged for 2.5 hr at 20,000 xg. The supernatant was removed and the pellet dissolved in 100 ml 8 M urea, 20 mM Tris-Cl pH 8.0, 5 mM DTT, 15 mM β -mercaptoethanol, 5 mM PMSF, and 1 mM KEDTA. The pellet was solubilized using a polytron homogenizer (Beckman, Berkeley, CA) and centrifuged at 20,000 xg for 2 hr.

3. DEAE chromatography:

Supernatant was loaded onto a 550 ml column (5 cm x 28 cm) packed with DEAE Fast Flow Sepharose (Pharmacia) equilibrated in 8 M urea, 20 mM Tris-Cl pH 8.0, 15 mM β -mercaptoethanol, and 1 mM KEDTA at room temperature. The column was washed with 1.5 liters equilibration buffer, and the protein eluted with a 5.0 liter linear gradient from 0-0.8 M NaCl in equilibration buffer. The HTLV-III protein eluted at 0.2 M NaCl and was assayed using SDS-polyacrylamide electrophoresis and following the protein at approximately 70 kD.

The fractions containing the HTLV-III protein were pooled and the protein concentrated to 10 ml using a stressed cell positive pressure concentrator

-34-

(Amicon) fitted with a 10,000 MW cut-off membrane (YM-10, Amicon). The concentrate was loaded onto a 500 ml column (2.5 cm x 100 cm) packed with superfine sephacryl S-300 (Pharmacia) equilibrated in 8 M urea, 20 mM Tris-Cl, pH 8.0, 15 mM β -mercaptoethanol, and 1 mM KEDTA. The column was eluted with equilibration buffer at room temperature. A flow rate of 0.5 ml/min was maintained. The HTLV-III protein eluted at approximately 40% of the column volume.

4. SDS-polyacrylamide electrophoresis:

The fractions containing KHI were pooled and the protein concentrated using a stressed cell positive pressure concentrator fitted with a 10,000 MW cutoff membrane. 2 mg of protein was mixed with loading buffers and electrophoresed through a preparative SDS-polyacrylamide gel (40 cm x 20 cm x 4 mm) as described by M.W. Hunkapiller, E. Lujan, F. Ostrander, and L.E. Hood, Methods in Enzymology 91:227-236 (1983). The 70 kD HTLV-III protein was visualized with Coomassie blue stain and eluted from the gel as described. The protein can be removed from the SDS by precipitation with acetone (Dyan, W.J., Jendrisak, J.J., Hager, D.A. and Burgess, R.R. [1981] J. Biol. Chem. 256:5860-5865).

-35-

Table 1

5' GATCAAGCTTCTGCAGTCGACGCATGCGGATCCGGTACCCGGGAGCTCG 3'
TTCGAAGACGTCAGCTGCGTACGCCTAGGCCATGGGCCCTCGAGCTTAA

Table 2

5' CGGTACCAGCCCGCCTAATGAGCGGGCTTTTTTTTGACGT 3'
TGCAGCCATGGTCGGGCGGATTACTCGCCCGAAAAAAAC

Table 3

MluI EcoRV ClaI BamHI Sali HindIII SmaI

CGAACGCGTGGCCGATATCATCGATGGATCCGTGACAAGCTTCCCGGGAGCT
GCTTGCGCACCGGCTATAGTAGCTACCTAGGCAGCTGTTTGAAGGGCCC

-36-

Table 4

5' AATTCCTGTGTGGAAGGAAGCA
TTAAGGGACACACCTTCCTTCGT

ACCACCACTCTATTTTGTGCATCAGATGCTAAAGCATATGATACAGAGGTACAT
TGGTGGTGAGATAAAACACGTAGTCTACGATTCGTATACTATGTCTCCATGTA

AATGTTTGGGCCACACATGCCTGTGTACCCACAGACCCCAACCCACAAGAAGTA
TTACAAACCCGGTGTGTACGGACACATGGGTGTCTGGGGTTGGGTGTTCTTCAT

GTATTGGTAAATGTGACAGAAAATTTTAACATGTGGAAAAATGACATGGTAGAA
CATAACCATTTACACTGTCTTTTAAATTTGTACACCTTTTACTGTACCATCTT

CAGATGCATGAGGATATAATCAGTTTATGGGATCAAAGCCTAAAGCCATGTGTA
GTCTACGTACTCCTATATTAGTCAAATACCCTAGTTTCGGATTCGGTACACAT

AAATTAACCCCACTCTGTGTTAGTTTAAAGTGCCTGATTTGAAGAATGATACT
TTTAATTGGGGTGAGACACAATCAAATTTACGTGACTAACTTCTTACTATGA

AATACCAATAGTAGTAGCGGGAGAATGATAATGGAGAAAGGAGAGATAAAAAAC
TTATGGTTATCATCATCGCCCTCTTACTATTACCTCTTTCCTCTCTATTTTTTG

TGCTCTTTCAATATCAGCACAAGCATAAGAGGTAAGGTGCAGAAAGAATATGCA
ACGAGAAAGTTATAGTCGTGTTTCGTATTCTCCATTCCACGTCTTTCCTTATACGT

TTTTTTTATAAACTTGATATAATACCAATAGATAATGATACTACCAGCTATACG
AAAAAATATTTGAACTATATTATGGTTATCTATTACTATGATGGTCGATATGC

TTGACAAGTTGTAACACCTCAGTCATTACACAGGCCTGTCCAAAGGTATCCTTT
AACTGTTCAACATTGTGGAGTCAGTAATGTGTCCGGACAGGTTTCCATAGGAAA

GAGCCAATTCCCATACATTATTGTGCCCCGGCTGGTTTTTGCGATTCTAAAATGT
CTCGGTTAAGGGTATGTAATAACACGGGGCCGACCAAACGCTAAGATTTTACA

ATAATAAGACGTTCAATGGAACAGGACCATGTACAAATGTCAGCACAGTACAA
TTATTATTCTGCAAGTTACCTTGTCCTGGTACATGTTTACAGTCGTGTCTATGTT

TGTACACATGGAATTAGGCCAGTAGTATCAACTCAACTGCTGTTAAATGGCAGT
ACATGTGTACCTTAATCCGGTCATCATAGTTGAGTTGACGACAATTTACCGTCA

CTAGCAGAAGAAGAGGTAGTAATTAGATCTGCCAATTTACAGACAATGCTAAA
GATCGTCTTCTTCTCCATCATTAATCTAGACGGTTAAAGTGTCTGTTACGATTT

ACCATAATAGTACAGCTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAAC
TGGTATTATCATGTGCGACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTTG

AACAATACAAGAAAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTGTGTT
TTGTTATGTTCTTTTTCATAGGCATAGGTCTCTCCTAATCCCTCTCGTAAACAA

-37-

Table 4 (cont.)

ACAATAGGAAAAATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCA
TGTTATCCTTTTTATCCTTTATACTCTGTTCTGTGAACATTGTAATCATCTCGT

AAATGGAATAACACTTTAAAACAGATAGATAGCAAATTAAGAGAACAATTTGGA
TTACCTTATTGTGAAATTTTGTCTATCTATCGTTTAATTCTCTTGTTAAACCT

AATAATAAAACAATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTA
TTATTATTTTGTATTAGAAATTCGTGAGGAGTCCTCCCCTGGGTCTTTAACAT

ACGCACAGTTTTAATTGTGGAGGGGAATTTTTCTACTGTAATTCAACACAACCTG
TGCGTGTCAAATTAACACCTCCCCTTAAAAAGATGACATTAAGTTGTGTTGAC

TTTAATAGTACTTGGTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACT
AAATTATCATGAACCAAATTATCATGAACCTCATGATTCCAGTTTATTGTGA

GAAGGAAGTGACACAATCACCCCTCCCATGCAGAATAAAACAAATTATAAACATG
CTTCCTTCACTGTGTTAGTGGGAGGGTACGTCTTATTTGTTTAATATTTGTAC

TGGCAGGAAGTAGGAAAAGCAATGTATGCCCCCTCCCATCAGTGGACAAATTAGA
ACCGTCCTTCATCCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTTAATCT

TGTTTCATCAAATATTACAGGGCTGCTATTAACAAGAGATGGTGGTAATAGCAAC
ACAAGTAGTTTATAATGTCCCGACGATAATTGTTCTCTACCACCATATCGTTG

AATGAGTCCGA
TTACTCAGGCTCTAG

3'

-38-

Table 5

5' CTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAAC
GACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTG

AACAATACAAGAAAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTGTGTT
TTGTTATGTTCTTTTTTCATAGGCATAGGTCTCTCCTGGTCCCTCTCGTAAACAA

ACAATAGGAAAAATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCA
TGTATCCTTTTTATCCTTTATACTCTGTTTCGTGTAACATTGTAATCATCTCGT

AAATGGAATAACACTTTTAAACAGATAGATAGCAAATTAAGAGAACAATTTGGA
TTACCTTATTGTGAAATTTTGTCTATCTATCGTTTAATTCTCTTGTAAACCT

AATAATAAAACAATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTA
TTATTATTTTGTATTAGAAATTCGTCAGGAGTCCTCCCCTGGGTCTTTAACAT

ACGCACAGTTTTTAATTGTGGAGGGGAATTTTTCTACTGTAATTCAACACAACCTG
TGCGTGTCAAATTAACACCTCCCCTTAAAAAGATGACATTAAGTTGTGTTGAC

TTTAATAGTACTTGGTTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACT
AAATTATCATGAACCAAATTATCATGAACCTCATGATTTCCCAGTTTATTGTGA

GAAGGAAGTGACACAATCACCTCCCATGCAGAATAAAACAAATTATAAACATG
CTTCCTTCACTGTGTTAGTGGGAGGGTACGTCTTATTTTGTTTAATATTTGTAC

TGGCAGGAAGTAGGAAAAGCAATGTATGCCCTCCCATCAGTGGACAAATTAGA
ACCGTCCTTCATCCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTTAATCT

TGTTTCATCAAATATTACAGGGCTGCTATTAACAAGAGATGGTGGTAATAGCAAC
ACAAGTAGTTTATAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTG

AATGAGTCCGA 3'
TTACTCAGGCTCTAG

Table 6

5' CTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAAC
GACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTTG

AACAATACAAGAAAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTGTGTT
TTGTTATGTTCTTTTTCATAGGCATAGGTCTCTCCTGGTCCCTCTCGTAAACAA

ACAATAGGAAAAATAGGAAATATGAGACAAGCACATTGTAAATTAGTAGAGCA
TGTTATCCTTTTATCCTTTATACTCTGTTTCGTGTAAACATTGTAATCATCTCGT

AAATGGAATAACACTTTTAAACAGATAGATAGCAAATTAAGAGAACAATTTGGA
TTTACCTTATTGTGAAATTTTGTCTATCTATCGTTTAATTCTCTTGTTAAACCT

AATAATAAAACAATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTA
TTATTATTTTGTATTAGAAATTCGTGAGGAGTCCTCCCCTGGGTCTTTAACAT

ACGCACAGTTTTAATTGTGGAGGGGAATTTTTCTACTGTAATTCAACACAACCTG
TGCGTGTCAAAATTAACACCTCCCCTTAAAAAGATGACATTAAGTTGTGTTGAC

TTTAATAGTACTTGGTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACT
AAATTATCATGAACCAAATTATCATGAACCTCATGATTTCCAGTTTATTGTGA

GAAGGAAGTGACACAATCACCTCCCATGCAGAATAAAACAAATTATAAACATG
CTTCCTTCACTGTGTTAGTGGGAGGGTACGTCTTATTTTGTTTAATATTGTAC

TGGCAGGAAGTAGGAAAAGCAATGTATGCCCCTCCCATCAGTGGACAAATTAGA
ACCGTCCTTCATCCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTAATCT

TGTTTCATCAAATATTACAGGGCTGCTATTAACAAGAGATGGTGGTAATAGCAAC
ACAAGTAGTTTATAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTG

AATGAGTCCGAGATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGA
TTACTCAGGCTCTAGAAGTCTGGACCTCCTCCTCTATACTCCCTGTTAACCTCT

AGTGAATTATATAAATATAAAGTAGTAAAAATTGAACCATTAGGAGTAGCACCC
TCACTTAATATATTTATATTTTCATCATTTTTTAACCTGGTAATCCTCATCGTGGG

ACCAAGGCAAAGAGAAGAGTGGTGCAGAGAGAAAAAGAGCAGTGGGAATAGGA
TGGTTCCGTTTCTCTTCTCACCACGTCTCTCTTTTTTCTCGTCACCCTTATCCT

GCTTTGTTCTTGGGTTCTTGGGAGCAGCAGGAAGCACTATGGGCGCAGCGTCA
CGAAACAAGGAACCCAAGAACCCTCGTCGTCTTCTGATACCCGCGTCGAGT

ATGACGCTGACGGTACAGGCCAGACAATTATTGTCTGGTATAGTGACGACGAG
TACTGCGACTGCCATGTCCGGTCTGTTAATAACAGACCATATCACGTCGTCGTC

AACAATTTGCTGAGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCACAGTC
TTGTTAAACGACTCCCGATAACTCCGCGTTGTCTGAGACAACGTTGAGTGTGAG

TGGGGCATCAAGCAGCTCCAGGCAAGAATCCTGGCTGTGGAAAGATACCTAAAG
ACCCCGTAGTTCGTGAGGTCCGTTCTTAGGACCGACACCTTTCTATGGATTC

Table 6 (cont..)

GATCAACAGCTCCTGGGGATTGTTGGGGTTGCTCTGGAAACTCATTTGCACCACT
CTAGTTGTCGAGGACCCCTAAACCCCAACGAGACCTTTTGAGTAAACGTGGTGA

GCTGTGCCTTGGAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTTGGAAT
CGACACGGAACCTTACGATCAACCTCATTATTTAGAGACCTTGTCTAAACCTTA

AACATGACCTGGATGGAGTGGGACAGAGAAATTAACAATTACACA 3'
TTGTACTGGACCTACCTCACCTGTCTCTTTAATTGTTAATGTGTTTCA

Table 7

5' AATTCCCTGTGTGGAAGGAAGCA
TTAAGGGACACACCTTCCTTCGT

ACCACCACTCTATTTTGTGCATCAGATGCTAAAGCATATGATACAGAGGTACAT
TGGTGGTGAGATAAAACACGTAGTCTACGATTTTCGTATACTATGTCTCCATGTA

AATGTTTGGGCCACACATGCCTGTGTACCCACAGACCCCAACCCACAAGAAGTA
TTACAAACCCGGTGTGTACGGACACATGGGTGTCTGGGGTTGGGTGTTCTTCAT

GTATTGGTAAATGTGACAGAAAATTTTAAACATGTGGAAAATGACATGGTAGAA
CATAACCATTTACACTGTCTTTTAAATTGTACACCTTTTACTGTACCATCTT

CAGATGCATGAGGATATAATCAGTTTATGGGATCAAAGCCTAAAGCCATGTGTA
GTCTACGTACTCCTATATTAGTCAAATACCTAGTTTCGGATTTCCGTACACAT

AAATTAACCCCACTCTGTGTTAGTTTAAAGTGCATGATTGAAGAATGATACT
TTTAATTGGGGTGAGACACAATCAAATTTACGTGACTAACTTCTTACTATGA

AATACCAATAGTAGTAGCGGGAGAATGATAATGGAGAAAGGAGAGATAAAAAAC
TTATGGTTATCATCATCGCCCTCTTACTATTACCTCTTTCCTCTCTATTTTTTG

TGCTCTTTCAATATCAGCACAAGCATAAGAGGTAAGGTGCAGAAAGAATATGCA
ACGAGAAAGTTATAGTCGTGTTTCGTATTCTCCATTCCACGTCCTTTCTTATACGT

TTTTTTTTATAAACTTGATATAATACCAATAGATAATGATACTACCAGCTATACG
AAAAAATATTTGAACTATATTATGGTTATCTATTACTATGATGGTGCATATGC

TTGACAAGTTGTAACACCTCAGTCATTACACAGGCCTGTCCAAAGGTATCCTTT
AACTGTTCAACATTGTGGAGTCAGTAATGTGTCCGGACAGGTTTCCATAGGAAA

GAGCCAATTCCCATACATTATTGTGCCCCGGCTGGTTTTGCGATTCTAAAATGT
CTCGGTTAAGGGTATGTAATAACACGGGGCCGACCAAAACGCTAAGATTTTACA

ATAATAAGACGTTCAATGGAACAGGACCATGTACAAATGTCAGCACAGTACAA
TTATTATTCTGCAAGTTACCTTGTCTCGGTACATGTTTACAGTCGTGTCATGTT

TGTACACATGGAATTAGGCCAGTAGTATCAACTCAACTGCTGTTAAATGGCAGT
ACATGTGTACCTTAATCCGGTCATCATAGTTGAGTTGACGACAATTTACCGTCA

CTAGCAGAAGAAGAGGTAGTAATTAGATCTGCCAATTTACAGACAATGCTAAA
GATCGTCTTCTTCTCCATCATTAACTAGACGGTTAAAGTGTCTGTTACGATTT

ACCATAATAGTACAGCTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAAC
TGGTATTATCATGTGCACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTTG

AACAATACAAGAAAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTGTGT
TTGTTATGTTCTTTTTCATAGGCATAGGTCTCTCCTAATCCCTCTCGTAAACAA

ACAATAGGAAAAATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCA
TGTTATCCTTTTTATCCTTTATACTCTGTTTCGTGTAACATTGTAATCATCTCGT

Table 7 (cont.)

AAATGGAATAACACTTTAAAACAGATAGATAGCAAATTAAGAGAACAAATTTGGA
TTTACCTTATTGTGAAATTTTGTCTATCTATCGTTAATTCTCTTGTTAAACCT

AATAATAAAACAATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTA
TTATTATTTTGTATTAGAAATTCGTCAGGAGTCCTCCCCTGGGTCTTTAACAT

ACGCACAGTTTTTAATTGTGGAGGGGAATTTTTCTACTGTAATTCAACACAACCTG
TGCGTGTCAAAATTAACACCTCCCCTTAAAAAGATGACATTAACTTGTGTTGAC

TTTAATAGTACTTGGTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACT
AAATTATCATGAACCAATTATCATGAACCTCATGATTTCCAGTTTATTGTGA

GAAGGAAGTGACACAATCACCTCCCATGCAGAATAAAACAAATTATAAACATG
CTTCCTTCACTGTGTTAGTGGGAGGGTACGTCTTATTTTGTTTAATATTTGTAC

TGGCAGGAAGTAGGAAAAGCAATGTATGCCCCTCCCATCAGTGGACAAATTAGA
ACCGTCCTTCATCCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTAATCT

TGTTTCATCAAATATTACAGGGCTGCTATTAACAAGAGATGGTGGTAATAGCAAC
ACAAGTAGTTTATAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTG

AATGAGTCCGAGATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGA
TTACTCAGGCTCTAGAAGTCTGGACCTCCTCCTCTATACTCCCTGTTAACCTCT

AGTGAATTATATAAATATAAAGTAGTAAAAATTGAACCATTAGGAGTAGCACCC
TCACTTAATATATTTATATTTTCATCATTTTTTAACTTGGTAATCCTCATCGTGGG

ACCAAGGCAAAGAGAAGAGTGGTGCAGAGAGAAAAAGAGCAGTGGGAATAGGA
TGGTTCCGTTTCTCTTCTCACCACGTCTCTCTTTTTTCTCGTCACCCCTTATCCT

GCTTTGTTCCCTTGGGTTCTTGGGAGCAGCAGGAAGCACTATGGGCGCAGCGTCA
CGAAACAAGGAACCCAAGAACCCTCGTCGTCTTTCGTGATACCCGCGTCGCAGT

ATGACGCTGACGGTACAGGCCAGACAATTATTGTCTGGTATAGTGCAGCAGCAG
TACTGCGACTGCCATGTCCGGTCTGTTAATAACAGACCATATCACGTCGTCGTC

AACAATTTGCTGAGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCACAGTC
TTGTTAAACGACTCCCGATAACTCCGCGTTGTCTGATAGACAACGTTGAGTGTGAG

TGGGGCATCAAGCAGCTCCAGGCAAGAATCCTGGCTGTGGAAAGATACCTAAAG
ACCCCGTAGTTCGTGAGGTCCGTTCTTAGGACCGACACCTTTCTATGGATTTC

GATCAACAGCTCCTGGGGATTTGGGGTTGCTCTGGAAAACCTCATTTGCACCACT
CTAGTTGTGAGGACCCCTAAACCCCAACGAGACCTTTTGAGTAAACGTGGTGA

GCTGTGCCTTGGAAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTTGGAAT
CGACACGGAACCTTACGATCAACCTCATTATTTAGAGACCTTGTCTAAACCTTA

AACATGACCTGGATGGAGTGGGACAGAGAAATTAACAATTACACA
TTGTACTGGACCTACCTCACCTGTCTCTTTAATTGTTAATGTGTTGCA

Table 8

Amino acid sequence of fusion protein R10

MetLeuArg
 ProValGluThrProThrArgGluIleLysLysLeuAspGlyLeuTrpAlaPhe
 SerLeuAspArgGluAsnCysGlyIleAspGlnPheProValTrpLysGluAla
 ThrThrThrLeuPheCysAlaSerAspAlaLysAlaTyrAspThrGluValHis
 AsnValTrpAlaThrHisAlaCysValProThrAspProAsnProGlnGluVal
 ValLeuValAsnValThrGluAsnPheAsnMetTrpLysAsnAspMetValGlu
 GlnMetHisGluAspIleIleSerLeuTrpAspGlnSerLeuLysProCysVal
 LysLeuThrProLeuCysValSerLeuLysCysThrAspLeuLysAsnAspThr
 AsnThrAsnSerSerSerGlyArgMetIleMetGluLysGlyGluIleLysAsn
 CysSerPheAsnIleSerThrSerIleArgGlyLysValGlnLysGluTyrAla
 PhePheTyrLysLeuAspIleIleProIleAspAsnAspThrThrSerTyrThr
 LeuThrSerCysAsnThrSerValIleThrGlnAlaCysProLysValSerPhe
 GluProIleProIleHisTyrCysAlaProAlaGlyPheAlaIleLeuLysCys
 AsnAsnLysThrPheAsnGlyThrGlyProCysThrAsnValSerThrValGln
 CysThrHisGlyIleArgProValValSerThrGlnLeuLeuLeuAsnGlySer
 LeuAlaGluGluGluValValIleArgSerAlaAsnPheThrAspAsnAlaLys
 ThrIleIleValGlnLeuAsnGlnSerValGluIleAsnCysThrArgProAsn
 AsnAsnThrArgLysSerIleArgIleGlnArgGlyProGlyArgAlaPheVal
 ThrIleGlyLysIleGlyAsnMetArgGlnAlaHisCysAsnIleSerArgAla
 LysTrpAsnAsnThrLeuLysGlnIleAspSerLysLeuArgGluGlnPheGly
 AsnAsnLysThrIleIlePheLysGlnSerSerGlyGlyAspProGluIleVal
 ThrHisSerPheAsnCysGlyGlyGluPhePheTyrCysAsnSerThrGlnLeu
 PheAsnSerThrTrpPheAsnSerThrTrpSerThrLysGlySerAsnAsnThr
 GluGlySerAspThrIleThrLeuProCysArgIleLysGlnIleIleAsnMet
 TrpGlnGluValGlyLysAlaMetTyrAlaProProIleSerGlyGlnIleArg
 CysSerSerAsnIleThrGlyLeuLeuLeuThrArgAspGlyGlyAsnSerAsn

Table 8 (cont.)

AsnGluSerGluIleHisArgSerValMetLeuTyrThrThrProAsnThrTrp
ValAspAspIleThrValValThrHisValAlaGlnAspCysAsnHisAlaSer
ValAspTrpGlnValValAlaAsnGlyAspValSerValGluLeuArgAspAla
AspGlnGlnValValAlaThrGlyGlnGlyThrSerGlyThrLeuGlnValVal
AsnProHisLeuTrpGlnProGlyGluGlyTyrLeuTyrGluLeuCysValThr
AlaLysSerGlnThrGluCysAspIleTyrProLeuArgValGlyIleArgSer
ValAlaValLysGlyGluGlnPheLeuIleAsnHisLysProPheTyrPheThr
GlyPheGlyArgHisGluAspAlaAspLeuArgGlyLysGlyPheAspAsnVal
LeuMetValHisAspHisAlaLeuMetAspTrpIleGlyAlaAsnSerTyrArg
ThrSerHisTyrProTyrAlaGluGluMetLeuAspTrpAlaAspGluHisGly
IleValValIleAspGluThrAlaAlaValGlyPheAsnLeuSerLeuGlyIle
GlyPheGluAlaGlyAsnLysProLysGluLeuTyrSerGluGluAlaValAsn
GlyGluThrGlnGlnAlaHisLeuGlnAlaIleLysGluLeuIleAlaArgAsp
LysAsnHisProSerValValMetTrpSerIleAlaAsnGluProAspThrArg
ProGlnGlyAlaArgGluTyrPheAlaProLeuAlaGluAlaThrArgLysLeu
AspProThrArgProIleThrCysValAsnValMetPheCysAspAlaHisThr
AspThrIleSerAspLeuPheAspValLeuCysLeuAsnArgTyrTyrGlyTrp
TyrValGlnSerGlyAspLeuGluThrAlaGluLysValLeuGluLysGluLeu
LeuAlaTrpGlnGluLysLeuHisGlnProIleIleIleThrGluTyrGlyVal
AspThrLeuAlaGlyLeuHisSerMetTyrThrAspMetTrpSerGluGluTyr
GlnCysAlaTrpLeuAspMetTyrHisArgValPheAspArgValSerAlaVal
ValGlyGluGlnValTrpAsnPheAlaAspPheAlaThrSerGlnGlyIleLeu
ArgValGlyGlyAsnLysLysGlyIlePheThrArgAspArgLysProLysSer
AlaAlaPheLeuLeuGlnLysArgTrpThrGlyMetAsnPheGlyGluLysPro
GlnGlnGlyGlyLysGln

Table 8A

Nucleotide sequence encoding fusion protein R10

ATGTTACGT
TACAATGCA

CCTGTAGAAACCCCAACCCGTGAAATCAAAAACTCGACGGCCTGTGGGCATTC
GGACATCTTTGGGGTTGGGCACTTTAGTTTTTTGAGCTGCCGGACACCCGTAAG
AGTCTGGATCGCGAAAACCTGTGGAATTGATCAATTCCTGTGTGGAAGGAAGCA
TCAGACCTAGCGCTTTTGACACCTTAAGGACACACCTTCCTTCGT
ACCACCACTCTATTTTGTGCATCAGATGCTAAAGCATATGATACAGAGGTACAT
TGGTGGTGAGATAAAACACGTAGTCTACGATTTTCGTATACTATGTCTCCATGTA
AATGTTTGGGCCACACATGCCTGTGTACCCACAGACCCCAACCCACAAGAAGTA
TTACAAACCCGGTGTGTACGGACACATGGGTGTCTGGGGTTGGGTGTTCTTCAT
GTATTGGTAAATGTGACAGAAAATTTTAACATGTGGAAAATGACATGGTAGAA
CATAACCATTTACACTGTCTTTTAAATTTGTACACCTTTTACTGTACCATCTT
CAGATGCATGAGGATATAATCAGTTTATGGGATCAAAGCCTAAAGCCATGTGTA
GTCTACGTACTCCTATATTAGTCAAATACCCTAGTTTCGGATTTTCGGTACACAT
AAATTAACCCCACTCTGTGTAGTTTAAAGTGCATGATTTGAAGAAATGATACT
TTAATTGGGGTGAGACACAATCAAATTTACGTGACTAAACTTCTTACTATGA
AATACCAATAGTAGTAGCGGGAGAATGATAATGGAGAAAGGAGAGATAAAAAAC
TTATGGTTATCATCATCGCCCTCTTACTATTACCTCTTTCCTCTCTATTTTTTG
TGCTCTTTCAATATCAGCACAAGCATAAGAGGTAAGGTGCAGAAAGAATATGCA
ACGAGAAAGTTATAGTCGTGTTTCTCCATTCCACGTCTTCTTATACGT
TTTTTTTATAAACTTGATATAATACCAATAGATAATGATACTACCAGCTATACG
AAAAAATATTTGAACTATATTATGGTTATCTATTACTATGATGGTCGATATGC
TTGACAAGTTGTAACACCTCAGTCATTACACAGGCCTGTCCAAAGGTATCCTTT
AACTGTTCAACATTGTGGAGTCAGTAATGTGTCCGGACAGGTTTCCATAGGAAA
GAGCCAATTCCCATACATTATTGTGCCCCGGCTGGTTTTGCGATTCTAAAATGT
CTCGGTTAAGGGTATGTAATAACACGGGGCCGACCAAAACGCTAAGATTTTACA
AATAATAAGACGTTCAATGGAACAGGACCATGTACAAATGTGAGCAGTACAA
TTATTATTCTGCAAGTTACCTTGTCTGGTACATGTTTACAGTCGTGTCATGTT
TGTAACATGGAATTAGGCCAGTAGTATCAACTCAACTGCTGTTAAATGGCAGT
ACATGTGTACCTTAATCCGGTCATCATAGTTGAGTTGACGACAATTTACCGTCA
CTAGCAGAAGAAGAGGTAGTAATTAGATCTGCCAATTTACAGACAATGCTAAA
GATCGTCTTCTTCTCCATCATTAATCTAGACGGTTAAAGTGTCTGTTACGATTT
ACCATAATAGTACAGCTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAAC
TGGTATTATCATGTGCGACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTTG

-46-

Table 8A (cont.)

AACAATACAAGAAAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTGTGTT
TTGTTATGTTCTTTTTTCATAGGCATAGGTCTCTCCTAATCCCTCTCGTAAACAA

ACAATAGGAAAAATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCA
TGTTATCCTTTTTATCCTTTATACTCTGTTCTGTGTAACATTGTAATCATCTCGT

AAATGGAATAACACTTTAAACAGATAGATAGCAAATTAAGAGAACAATTTGGA
TTTACCTTATTGTGAAATTTTGTCTATCTATCGTTTAATTCTCTTGTTAAACCT

AATAATAAAACAATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTA
TTATTATTTGTTATTAGAAATTCGTGAGGAGTCCTCCCCTGGGTCTTTAACAT

ACGCACAGTTTTTAATTGTGGAGGGGAATTTTTCTACTGTAATTCAACACAACCTG
TGCGTGTCAAATTAACACCTCCCCTTAAAAAGATGACATTAAGTTGTGTTGAC

TTTAATAGTACTTGGTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACT
AAATTATCATGAACCAAATTATCATGAACCTCATGATTTCCAGTTTATTGTGA

GAAGGAAGTGACACAATCACCTCCCATGCAGAATAAAACAAATTATAAACATG
CTTCCTTCACTGTGTTAGTGGGAGGGTACGTCTTATTTTGTTAATATTTGTAC

TGGCAGGAAGTAGGAAAAGCAATGTATGCCCTCCCATCAGTGGACAAATTAGA
ACCGTCCTTCATCCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTAATCT

TGTTTCATCAAATATTACAGGGCTGCTATTAACAAGAGATGGTGGTAATAGCAAC
ACAAGTAGTTTATAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTG

AATGAGTCCGAGATCCATCGCAGCGTAATGCTCTACACCACGCCGAACACCTGG
TTACTCAGGCTCTAGGTAGCGTCGCATTACGAGATGTGGTGCGGCTTGTGGACC

GTGGACGATATCACCGTGGTGACGCATGTCGCGCAAGACTGTAACCACGCGTCT
CACCTGCTATAGTGGCACCCTGCGTACAGCGCGTTCTGACATTGGTGCGCAGA

GTTGACTGGCAGGTGGTGGCCAATGGTGATGTCAGCGTTGAACTGCGTGATGCG
CAACTGACCGTCCACCACCGGTTACCACTACAGTCGCAACTTGACGCACTACGC

GATCAACAGGTGGTTGCAACTGGACAAGGCACTAGCGGGACTTTGCAAGTGGTG
CTAGTTGTCCACCAACGTTGACCTGTTCCGTGATCGCCCTGAAACGTTACCAC

AATCCGCACCTCTGGCAACCGGGTGAAGGTTATCTCTATGAACTGTGCGTCACA
TTAGGCGTGGAGACCGTTGGCCCACTTCCAATAGAGATACTTGACACGCAGTGT

GCCAAAAGCCAGACAGAGTGTGATATCTACCCGCTTCGCGTCGGCATCCGGTCA
CGGTTTTCGGTCTGTCTCACACTATAGATGGGCGAAGCGCAGCCGTAGGCCAGT

GTGGCAGTGAAGGGCGAACAGTTCCTGATTAACCACAAACCGTTCTACTTTACT
CACCGTCACTTCCCGCTTGTCAGGACTAATTGGTGTGTTGGCAAGATGAAATGA

GGCTTTGGTCGTTCATGAAGATGCGGACTTGCGTGGCAAAGGATTCGATAACGTG
CCGAAACCAGCAGCACTTCTACGCCTGAACGCACCGTTTCCCTAAGCTATTGCAC

CTGATGGTGCACGACCACGCATTAATGGACTGGATTGGGGCCAACTCCTACCGT
GACTACCACGTGCTGGTGCGTAATTACCTGACCTAACCCCGGTTGAGGATGGCA

Table 8A (cont.)

ACCTCGCATTACCCTTACGCTGAAGAGATGCTCGACTGGGCAGATGAACATGGC
TGGAGCGTAATGGGAATGCGACTTCTCTACGAGCTGACCCGTCTACTTGTACCG

ATCGTGGTGATTGATGAAACTGCTGCTGTGCGGCTTTAACCTCTCTTTAGGCATT
TAGCACCCTAACTACTTTGACGACGACAGCCGAAATTGGAGAGAAATCCGTAA

GGTTTCGAAGCGGGCAACAAGCCGAAAGAACTGTACAGCGAAGAGGCAGTCAAC
CCAAAGCTTCGCCCGTTGTTGCGGCTTTCTTGACATGTCGCTTCTCCGTCAGTTG

GGGGAACTCAGCAAGCGCACTTACAGGCGATTAAAGAGCTGATAGCGCGTGAC
CCCCTTTGAGTCGTTGCGGTGAATGTCCGCTAATTTCTCGACTATCGCGCACTG

AAAAACCACCCAAGCGTGGTGATGTGGAGTATTGCCAACGAACCGGATACCCGT
TTTTTGGTGGGTTGCGCACCCTACACCTCATAACGGTTGCTTGGCCTATGGGCA

CCGCAAGGTGCACGGGAATATTTGCGGCCACTGGCGGAAGCAACGCGTAAACTC
GGCGTTCCACGTGCCCTTATAAAGCGCGGTGACCGCTTCGTTGCGCATTGTAG

GACCCGACGCGTCCGATCACCTGCGTCAATGTAATGTTCTGCGACGCTCACACC
CTGGGCTGCGCAGGCTAGTGGACGCAGTTACATTACAAGACGCTGCGAGTGTGG

GATACCATCAGCGATCTCTTTGATGTGCTGTGCCTGAACCGTTATTACGGATGG
CTATGGTAGTCGCTAGAGAACTACACGACACGGACTTGGCAATAATGCCTACC

TATGTCCAAAGCGGCGATTTGGAAACGGCAGAGAAGGTACTGGAAAAAGAACTT
ATACAGGTTTCGCCGCTAAACCTTTGCCGTCTCTTCCATGACCTTTTCTTGAA

CTGGCCTGGCAGGAGAACTGCATCAGCCGATTATCATCACCGAATACGGCGTG
GACCGGACCGTCCTCTTTGACGTAGTCGGCTAATAGTAGTGGCTTATGCCGCAC

GATACGTTAGCCGGGCTGCACTCAATGTACACCGACATGTGGAGTGAAGAGTAT
CTATGCAATCGGCCCGACGTGAGTTACATGTGGCTGTACACCTCACTTCTCATA

CAGTGTGCATGGCTGGATATGTATCACCGCGTCTTTGATCGCGTCAGCGCCGTC
GTCACACGTACCGACCTATACATAGTGGCGCAGAACTAGCGCAGTCGCGGCAG

GTCGGTGAACAGGTATGGAATTTGCCGATTTTGGCAGCTCGCAAGGCATATTG
CAGCCACTTGTCCATACCTTAAAGCGGCTAAAACGCTGGAGCGTTCCGTATAAC

CGCGTTGGCGGTAAACAAGAAAGGGATCTTCACTCGCGACCGCAAACCGAAGTCG
GCGCAACCGCCATTGTTCTTTCCCTAGAAGTGAGCGCTGGCGTTTGGCTTCAGC

GCGGCTTTTCTGCTGCAAAAACGCTGGACTGGCATGAACTTCGGTGAAAAACCG
CGCCGAAAAGACGACGTTTTTGGCAGCTGACCGTACTTGAAGCCACTTTTTTGGC

CAGCAGGGAGGCAAACAA
GTCGTCCCTCCGTTTGT

Table 9

Amino acid sequence of fusion protein PBl

MetLeuArg

ProValGluThrProThrArgGluIleLysLysLeuAspGlyLeuTrpAlaPhe
SerLeuAspArgGluArgValAlaAspLeuAsnGlnSerValGluIleAsnCys
ThrArgProAsnAsnAsnThrArgLysSerIleArgIleGlnArgGlyProGly
ArgAlaPheValThrIleGlyLysIleGlyAsnMetArgGlnAlaHisCysAsn
IleSerArgAlaLysTrpAsnAsnThrLeuLysGlnIleAspSerLysLeuArg
GluGlnPheGlyAsnAsnLysThrIleIlePheLysGlnSerSerGlyGlyAsp
ProGluIleValThrHisSerPheAsnCysGlyGlyGluPhePheTyrCysAsn
SerThrGlnLeuPheAsnSerThrTrpPheAsnSerThrTrpSerThrLysGly
SerAsnAsnThrGluGlySerAspThrIleThrLeuProCysArgIleLysGln
IleIleAsnMetTrpGlnGluValGlyLysAlaMetTyrAlaProProIleSer
GlyGlnIleArgCysSerSerAsnIleThrGlyLeuLeuLeuThrArgAspGly
GlyAsnSerAsnAsnGluSerGluIleArgArgGlnAlaSerArgGluLeuGlu
PheLeuLysThrLysGlyProArgAspThrProIlePheIleGly

Table 9A

Nucleotide sequence encoding fusion protein PB1

ATGTTACGTCCTGTAGAAACCCCAACCCGTGAAATCAAAAACTCGACGGCCTG
TACAATGCAGGACATCTTTGGGGTTGGGCACCTTAGTTTTTTGAGCTGCCGGAC
TGGGCATTTCAGTCTGGATCGCGAACGCGTGGCCGATCTGAACCAATCTGTAGAA
ACCCGTAAGTCAGACCTAGCGCTTGCGCACCGGCTAGACTTGGTTAGACATCTT
ATTAATTGTACAAGACCCCAACAACAATACAAGAAAAAGTATCCGTATCCAGAGA
TAATTAACATGTTCTGGGTGTTGTTATGTTCTTTTTCATAGGCATAGGTCTCT
GGACCAGGGAGAGCATTGTTACAATAGGAAAAATAGGAAATATGAGACAAGCA
CCTAATCCCTCTCGTAAACAATGTTATCCTTTTTATCCTTTATACTCTGTTCTG
CATTGTAACATTAGTAGAGCAAAATGGAATAACACTTTAAACAGATAGATAGC
GTAACATTGTAATCATCTCGTTTTACCTTATTGTGAAATTTTGTCTATCTATCG
AAATTAAGAGAACAATTTGGAAATAATAAAACAATAATCTTTAAGCAGTCCTCA
TTTAATTCTCTTGTTAAACCTTTATTATTTTGTATTAGAAATTCTGCAGGAGT
GGAGGGGACCCAGAAATTGTAACGCACAGTTTTAATTGTGGAGGGGAATTTTTTC
CCTCCCCTGGGTCTTTAACATTGCGTGTCAAATTAACACCTCCGCTTAAAAAG
TACTGTAATTCAACACAACCTGTTTAATAGTACTTGGTTTAATAGTACTTGGAGT
ATGACATTAAAGTTGTGTTGACAAATTATCATGAACCAAATTATCATGAACCTCA
ACTAAAGGGTCAAATAACACTGAAGGAAGTGACACAATCACCCCTCCCATGCGAGA
TGATTTCCAGTTTATTGTGACTTCCTTCACTGTGTTAGTGGGAGGGTACGTCT
ATAAAACAAATTATAAACATGTGGCAGGAAGTAGGAAAAGCAATGTATGCCCT
TATTTTGTTTAATATTTGTACACCGTCCTTCATCCTTTTCGTTACATACGGGGA
CCCATCAGTGGACAAATTAGATGTTTCATCAAATATTACAGGGCTGCTATTAAACA
GGGTAGTCACCTGTTTAATCTACAAGTAGTTTATAATGTCCCGACGATAATTGT
AGAGATGGTGGTAATAGCAACAATGAGTCCGAGATCCGTCGACAAGCTTCCCGG
TCTCTACCACATTATCGTTGTTACTCAGGCTCTAGGCAGCTGTTCTGAAGGGCC
GAGCTCGAATTCTTGAAGACGAAAGGGCCTCGTGATACTCCTATTTTTATAGGT
CTCGAGCTTAAGAACTTCTGCTTTCCCGGAGCACTATGCGGATAAAAATATCCA

Table 10

Amino acid sequence of fusion protein 590

MetLeuArgProValGluThr
 ProThrArgGluIleLysLysLeuAspGlyLeuTrpAlaPheSerLeuAspArg
 GluArgValAlaAspLeuAsnGlnSerValGluIleAsnCysThrArgProAsn
 AsnAsnThrArgLysSerIleArgIleGlnArgGlyProGlyArgAlaPheVal
 ThrIleGlyLysIleGlyAsnMetArgGlnAlaHisCysAsnIleSerArgAla
 LysTrpAsnAsnThrLeuLysGlnIleAspSerLysLeuArgGluGlnPheGly
 AsnAsnLysThrIleIlePheLysGlnSerSerGlyGlyAspProGluIleVal
 ThrHisSerPheAsnCysGlyGlyGluPhePheTyrCysAsnSerThrGlnLeu
 PheAsnSerThrTrpPheAsnSerThrTrpSerThrLysGlySerAsnAsnThr
 GluGlySerAspThrIleThrLeuProCysArgIleLysGlnIleIleAsnMet
 TrpGlnGluValGlyLysAlaMetTyrAlaProProIleSerGlyGlnIleArg
 CysSerSerAsnIleThrGlyLeuLeuLeuThrArgAspGlyGlyAsnSerAsn
 AsnGluSerGluIlePheArgProGlyGlyGlyAspMetArgAspAsnTrpArg
 SerGluLeuTyrLysTyrLysValValLysIleGluProLeuGlyValAlaPro
 ThrLysAlaLysArgArgValValGlnArgGluLysArgAlaValGlyIleGly
 AlaLeuPheLeuGlyPheLeuGlyAlaAlaGlySerThrMetGlyAlaAlaSer
 MetThrLeuThrValGlnAlaArgGlnLeuLeuSerGlyIleValGlnGlnGln
 AsnAsnLeuLeuArgAlaIleGluAlaGlnGlnHisLeuLeuGlnLeuThrVal
 TrpGlyIleLysGlnLeuGlnAlaArgIleLeuAlaValGluArgTyrLeuLys
 AspGlnGlnLeuLeuGlyIleTrpGlyCysSerGlyLysLeuIleCysThrThr
 AlaValProTrpAsnAlaSerTrpSerAsnLysSerLeuGluGlnIleTrpAsn
 AsnMetThrTrpMetGluTrpAspArgGluIleAsnAsnTyrThrSerPhePro
 IleHisArgSerValMetLeuTyrThrThrProAsnThrTrpValAspAspIle
 ThrValValThrHisValAlaGlnAspCysAsnHisAlaSerValAspTrpGln
 ValValAlaAsnGlyAspValSerValGluLeuArgAspAlaAspGlnGlnVal

Table 10 (cont.)

ValAlaThrGlyGlnGlyThrSerGlyThrLeuGlnValValAsnProHisLeu
TrpGlnProGlyGluGlyTyrLeuTyrGluLeuCysValThrAlaLysSerGln
ThrGluCysAspIleTyrProLeuArgValGlyIleArgSerValAlaValLys
GlyGluGlnPheLeuIleAsnHisLysProPheTyrPheThrGlyPheGlyArg
HisGluAspAlaAspLeuArgGlyLysGlyPheAspAsnValLeuMetValHis
AspHisAlaLeuMetAspTrpIleGlyAlaAsnSerTyrArgThrSerHisTyr
ProTyrAlaGluGluMetLeuAspTrpAlaAspGluHisGlyIleValValIle
AspGluThrAlaAlaValGlyPheAsnLeuSerLeuGlyIleGlyPheGluAla
GlyAsnLysProLysGluLeuTyrSerGluGluAlaValAsnGlyGluThrGln
GlnAlaHisLeuGlnAlaIleLysGluLeuIleAlaArgAspLysAsnHisPro
SerValValMetTrpSerIleAlaAsnGluProAspThrArgProGlnGlyAla
ArgGluTyrPheAlaProLeuAlaGluAlaThrArgLysLeuAspProThrArg
ProIleThrCysValAsnValMetPheCysAspAlaHisThrAspThrIleSer
AspLeuPheAspValLeuCysLeuAsnArgTyrTyrGlyTrpTyrValGlnSer
GlyAspLeuGluThrAlaGluLysValLeuGluLysGluLeuLeuAlaTrpGln
GluLysLeuHisGlnProIleIleIleThrGluTyrGlyValAspThrLeuAla
GlyLeuHisSerMetTyrThrAspMetTrpSerGluGluTyrGlnCysAlaTrp
LeuAspMetTyrHisArgValPheAspArgValSerAlaValValGlyGluGln
ValTrpAsnPheAlaAspPheAlaThrSerGlnGlyIleLeuArgValGlyGly
AsnLysLysGlyIlePheThrArgAspArgLysProLysSerAlaAlaPheLeu
LeuGlnLysArgTrpThrGlyMetAsnPheGlyGluLysProGlnGlnGlyGly
LysGln

-52-

Table 10A

Nucleotide sequence encoding fusion protein 590

ATGTTACGTCCTGTAGAAACC
TACAATGCAGGACATCTTTGG

CCAACCCGTGAAATCAAAAACTCGACGGCCTGTGGGCATTGAGTCTGGATCGC
GGTTGGGCACTTTAGTTTTTTGAGATGCCGGACACCCGTAAGTCAGACCTAGCG

GAACGCGTGGCCGATCTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAAC
CTTGCGCACCGGCTAGACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTTG

AACAATACAAGAAAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTGTGTT
TTGTTATGTTCTTTTTCATAGGCATAGGTCTCTCCTAATCCCTCTCGTAAACAA

ACAATAGGAAAAATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCA
TGTTATCCTTTTTATCCTTTATACTCTGTTTCGTGTAACATTGTAATCATCTCGT

AAATGGAATAACACTTTAAAACAGATAGATAGCAAATTAAGAGAACAATTTGGA
TTTACCTTATTGTGAAATTTTGTCTATCTATCGTTTAATTCTCTTGTTAAACCT

AATAATAAAACAATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTA
TTATTATTTTGTTATTAGAAATTCGTCAGGAGTCCTCCCCTGGGTCTTTAACAT

ACGCACAGTTTTAATTGTGGAGGGGAATTTTTCTACTGTAATTCAACACAACCTG
TGCGTGTCAAATTAACACCTCCCCTTAAAAAGATGACATTAAGTTGTGTTGAC

TTTAATAGTACTTGGTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACT
AAATTATCATGAACCAAATTATCATGAACCTCATGATTCCCAGTTTATTGTGA

GAAGGAAGTGACACAATCACCCCTCCCATGCAGAATAAAACAAATTATAAACATG
CTTCCTTCACCTGTGTTAGTGGGAGGGTACGTCTTATTTTGTTTAATATTGTAC

TGGCAGGAAGTAGGAAAAGCAATGTATGCCCCCTCCCATCAGTGGACAAATTAGA
ACCGTCCTTCATCCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTTAATCT

TGTTTCATCAAATATTACAGGGCTGCTATTAACAAGAGATGGTGGTAATAGCAAC
ACAAGTAGTTTATAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTG

AATGAGTCCGAGATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGA
TTACTCAGGCTCTAGAAGTCTGGACCTCCTCCTCTATACTCCCTGTTAACCTCT

AGTGAATTATATAAAATATAAAGTAGTAAAAAATTGAACCATTAGGAGTAGCACCC
TCACTTAATATATTTATATTTTCATCATTTTTTAACCTTGGTAATCCTCATCGTGGG

ACCAAGGCAAAGAGAAGAGTGGTGCAGAGAGAAAAAAGAGCAGTGGGAATAGGA
TGGTCCGTTTTCTCTCTCACCACGTCTCTCTTTTTTCTCGTCACCCCTATCCT

Table 10A (cont.)

GCTTTGTTCTTGGGTTCTTGGGAGCAGCAGGAAGCACTATGGGCGCAGCGTCA
CGAAACAAGGAACCCAAGAACCCTCGTCGTCCTTCGTGATACCCGCGTCGAGT
ATGACGCTGACGGTACAGGCCAGACAATTATTGTCTGGTATAGTGACAGCAGCAG
TACTGCGACTGCCATGTCCGGTCTGTTAATAACAGACCATATCACGTCGTCGTC
AACAAATTTGCTGAGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCACAGTC
TTGTTAAACGACTCCCATAACTCCGCGTTGTCGTAGACAACGTTGAGTGTACG
TGGGGCATCAAGCAGCTCCAGGCAAGAATCCTGGCTGTGGAAAGATACCTAAAG
ACCCCGTAGTTCGTGAGGTCCGTTCTTAGGACCGACACCTTTCTATGGATTTC
GATCAACAGCTCCTGGGGATTTGGGGTTGCTCTGGAAAACCTATTGACCACT
CTAGTTGTGAGGACCCCTAAACCCCAACGAGACCTTTTGAGTAAACGTGGTGA
GCTGTGCCTTGAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTTGGAAT
CGACACGGAACCTTACGATCAACCTCATTATTTAGAGACCTTGCTAAACCTTA
AACATGACCTGGATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTCCCG
TTGTACTGGACCTACCTCACCTGTCTCTTTAATTGTTAATGTGPTCGAAGGGC
ATCCATCGCAGCGTAATGCTCTACACCACGCCGAACACCTGGGTGGACGATATC
TAGGTAGCGTCGCATTACGAGATGTGGTGCGGCTTGTGGACCCACCTGCTATAG
ACCGTGGTGACGCATGTGCGGCAAGACTGTAACCACGCGTCTGTTGACTGGCAG
TGGCACCCTGCGTACAGCGCGTTCTGACATTGGTGCGCAGACAACCTGACCGTC
GTGGTGGCCAATGGTGATGTGAGCGTTGAACTGCGTGATGCGGATCAACAGGTG
CACCACCGGTTACCACTACAGTCGCAACTTGACGCACTACGCTAGTTGTCCAC
GTTGCAACTGGACAAGGCACTAGCGGGACTTTGCAAGTGGTGAATCCGCACCTC
CAACGTTGACCTGTTCCGTGATCGCCCTGAAACGTTACCACTTAGGCGTGGAG
TGGCAACCGGGTGAAGGTTATCTCTATGAACTGTGCGTCACAGCCAAAAGCCAG
ACCGTTGGCCCACTTCCAATAGAGATACTTGACACGCAGTGTGCGTTTTCGGTC
ACAGAGTGTGATATCTACCCGCTTCGCGTCGGCATCCGGTCAGTGGCAGTGAAG
TGTCTCACACTATAGATGGGCGAAGCGCAGCCGTAGGCCAGTCACCGTCACTTC
GGCGAACAGTTCCTGATTAACCACAAACCGTTCTACTTTACTGGCTTTGGTCGT
CCGCTTGTCAAGGACTAATTGGTGTGTTGGCAAGATGAAATGACCGAAACCAGCA
CATGAAGATGCGGACTTGCGTGCGCAAGGATTGATAACGTGCTGATGGTGCAC
GCACTTCTACGCCTGAACGCACCGTTTCCTAAGCTATTGCACGACTACCACGTG
GACCACGCATTAATGGACTGGATTGGGGCCAACTCCTACCGTACCTCGCATTAC
CTGGTGCGTAATTACCTGACCTAACCCCGGTTGAGGATGGCATGGAGCGTAATG

Table 10A (cont.)

CCTTACGCTGAAGAGATGCTCGACTGGGCAGATGAACATGGCATCGTGGTGATT
GGAATGCGACTTCTCTACGAGCTGACCCGTCTACTTGTACCGTAGCACCCTAA

GATGAACTGCTGCTGTCTGGCTTTAACCTCTCTTTAGGCATTGGTTTCGAAGCG
CTACTTTGACGACGACAGCCGAAATTGGAGAGAAATCCGTAACCAAAGCTTCGC

GGCAACAAGCCGAAAGAACTGTACAGCGAAGAGGCAGTCAACGGGGAAACTCAG
CCGTTGTTCTGGCTTTCTTGACATGTCGCTTCTCCGTCAGTTGCCCCCTTGAGTC

CAAGCGCACTTACAGGCGATTAAAGAGCTGATAGCGCGTGACAAAAACCACCCA
GTTGCGGTGAATGTCCGCTAATTTCTCGACTATCGCGCACTGTTTTTGGTGGGT

AGCGTGGTGATGTGGAGTATTGCCAACGAACCGGATACCCGTCCGCAAGGTGCA
TCGCACCACTACACCTCATAACGGTTGCTTGGCCTATGGGCAGGCGTTCACGT

CGGGAATATTTCTCGGCCACTGGCGGAAGCAACGCGTAAACTCGACCCGACGCGT
GCCCTTATAAAGCGCGGTGACCGCCTTCGTTGCGCATTTGAGCTGGGCTGCGCA

CCGATCACCTGCGTCAATGTAATGTTCTGCGACGCTCACACCGATACCATCAGC
GGCTAGTGGACGCAGTTACATTACAAGACGCTGCGAGTGTGGCTATGGTAGTCG

GATCTCTTTGATGTGCTGTGCCTGAACCGTTATTACGGATGGTATGTCCAAAGC
CTAGAGAACTACACGACACGGACTTGGCAATAATGCCTACCATACAGGTTTCG

GGCGATTTGGAAACGGCAGAGAAGGTACTGGAAAAAGAACTTCTGGCCTGGCAG
CCGCTAAACCTTTGCCGTCTCTTCCATGACCTTTTTCTTGAAGACCGGACCGTC

GAGAACTGCATCAGCCGATTATCATCACCGAATACGGCGTGGATACGTTAGCC
CTCTTTGACGTAGTCGGCTAATAGTAGTGGCTTATGCCGCACCTATGCAATCGG

GGGCTGCACTCAATGTACACCGACATGTGGAGTGAAGAGTATCAGTGTGCATGG
CCCGACGTGAGTTACATGTGGCTGTACACCTCACTTCTCATAGTCACACGTACC

CTGGATATGTATCACCGCGTCTTTGATCGCGTCAGCGCCGTGCTCGGTGAACAG
GACCTATACATAGTGGCGCAGAACTAGCGCAGTGC CGGCAGCAGCCACTTGTC

GTATGGAATTTGCGCGATTTTGGCGACCTCGCAAGGCATATTGCGCGTTGGCGGT
CATACCTTAAAGCGGCTAAAACGCTGGAGCGTTCCGTATAACGCGCAACCGCCA

AACAAGAAAGGGATCTTCACTCGCGACCGCAAACCGAAGTCGGCGGCTTTTCTG
TTGTTCTTTCCCTAGAAGTGAGCGCTGGCGTTTGGCTTCAGCCGCCGAAAAGAC

CTGCAAAAACGCTGGACTGGCATGAACTTCGGTGAAAAACCGCAGCAGGGAGGC
GACGTTTTTGGCGACCTGACCGTACTTGAAGCCACTTTTTGGCGTCGTCCCTCCG

AAACAA
TTTGTT

Table 11
Amino acid sequence of fusion protein KH1

MetLeuArg
ProValGluThrProThrArgGluIleLysLysLeuAspGlyLeuTrpAlaPhe
SerLeuAspArgGluArgGluPheProValTrpLysGluAlaThrThrThrLeu
PheCysAlaSerAspAlaLysAlaTyrAspThrGluValHisAsnValTrpAla
ThrHisAlaCysValProThrAspProAsnProGlnGluValValLeuValAsn
ValThrGluAsnPheAsnMetTrpLysAsnAspMetValGluGlnMetHisGlu
AspIleIleSerLeuTrpAspGlnSerLeuLysProCysValLysLeuThrPro
LeuCysValSerLeuLysCysThrAspLeuLysAsnAspThrAsnThrAsnSer
SerSerGlyArgMetIleMetGluLysGlyGluIleLysAsnCysSerPheAsn
IleSerThrSerIleArgGlyLysValGlnLysGluTyrAlaPhePheTyrLys
LeuAspIleIleProIleAspAsnAspThrThrSerTyrThrLeuThrSerCys
AsnThrSerValIleThrGlnAlaCysProLysValSerPheGluProIlePro
IleHisTyrCysAlaProAlaGlyPheAlaIleLeuLysCysAsnAsnLysThr
PheAsnGlyThrGlyProCysThrAsnValSerThrValGlnCysThrHisGly
IleArgProValValSerThrGlnLeuLeuLeuAsnGlySerLeuAlaGluGlu
GluValValIleArgSerAlaAsnPheThrAspAsnAlaLysThrIleIleVal
GlnLeuAsnGlnSerValGluIleAsnCysThrArgProAsnAsnAsnThrArg
LysSerIleArgIleGlnArgGlyProGlyArgAlaPheValThrIleGlyLys
IleGlyAsnMetArgGlnAlaHisCysAsnIleSerArgAlaLysTrpAsnAsn
ThrLeuLysGlnIleAspSerLysLeuArgGluGlnPheGlyAsnAsnLysThr
IleIlePheLysGlnSerSerGlyGlyAspProGluIleValThrHisSerPhe
AsnCysGlyGlyGluPhePheTyrCysAsnSerThrGlnLeuPheAsnSerThr
TrpPheAsnSerThrTrpSerThrLysGlySerAsnAsnThrGluGlySerAsp
ThrIleThrLeuProCysArgIleLysGlnIleIleAsnMetTrpGlnGluVal

Table 11 (cont.)

GlyLysAlaMetTyrAlaProProIleSerGlyGlnIleArgCysSerSerAsn
IleThrGlyLeuLeuLeuThrArgAspGlyGlyAsnSerAsnAsnGluSerGlu
IlePheArgProGlyGlyGlyAspMetArgAspAsnTrpArgSerGluLeuTyr
LysTyrLysValValLysIleGluProLeuGlyValAlaProThrLysAlaLys
ArgArgValValGlnArgGluLysArgAlaValGlyIleGlyAlaLeuPheLeu
GlyPheLeuGlyAlaAlaGlySerThrMetGlyAlaAlaSerMetThrLeuThr
ValGlnAlaArgGlnLeuLeuSerGlyIleValGlnGlnGlnAsnAsnLeuLeu
ArgAlaIleGluAlaGlnGlnHisLeuLeuGlnLeuThrValTrpGlyIleLys
GlnLeuGlnAlaArgIleLeuAlaValGluArgTyrLeuLysAspGlnGlnLeu
LeuGlyIleTrpGlyCysSerGlyLysLeuIleCysThrThrAlaValProTrp
AsnAlaSerTrpSerAsnLysSerLeuGluGlnIleTrpAsnAsnMetThrTrp
MetGluTrpAspArgGluIleAsnAsnTyrThrSerPheProGlyAlaArgIle
LeuGluAspGluArgAlaSer

Table 11A

Nucleotide sequence encoding fusion protein KH1

ATGTTACGT
TACAATGCACCTGTAGAAACCCCAACCCGTGAAATCAAAAACTCGACGGCCTGTGGGCATTC
GGACATCTTTGGGGTTGGGCACCTTTAGTTTTTTGAGCTGCCGGACACCCGTAAGAGTCTGGATCGCGAACGCGAATTCCTGTGTGGAAGGAAGCAACCACCACTCTA
TCAGACCTAGCGCTTGCGCTTAAGGGACACACCTTCCTTCGTTGGTGGTGGATTTTTGTGCATCAGATGCTAAAGCATATGATACAGAGGTACATAATGTTTGGGCC
AAAACACGTAGTCTACGATTTCTGTATACTATGTCTCCATGTATTACAAACCCGGACACATGCCTGTGTACCCACAGACCCCAACCCACAAGAAAGTATGTTAAAT
TGTGTACGGACACATGGGTGTCTGGGGTTGGGTGTTCTTCATCATAACCATTTAGTGACAGAAAATTTTAAACATGTGGAATAATGACATGGTAGAACAGATGCATGAG
CACTGTCTTTTAAATTTGTACACCTTTTTACTGTACCATCTTGTCTACGTACTCGATATAATCAGTTTATGGGATCAAAGCCTAAAGCCATGTGTAAAATTAACCCCA
CTATATTAGTCAAATACCCTAGTTTCGGATTTCGGTACACATTTTAATTGGGGTCTCTGTGTTAGTTTAAAGTGCACTGATTTGAAGAATGATACTAATACCAATAGT
GAGACACAATCAAATTTACGTGACTAAACTTCTTACTATGATTATGGTTATCAAGTAGCGGGAGAATGATAATGGAGAAAGGAGAGATAAAAACTGCTCTTTCAAT
TCATCGCCCTCTTACTATTACCTCTTCTCTATTTTTTTGACGAGAAAGTTAATCAGCACAAGCATAAGAGGTAAGGTGCAGAAAGAATATGCATTTTTTTTATAAA
TAGTCGTGTTTCGTATTCTCCATTCCACGTCTTCTTATACGTAAAAAATATTTCTTGATATAATACCAATAGATAATGATACTACCAGCTATACGTTGACAAGTTGT
GAACTATATTATGGTTATCTATTACTATGATGGTCGATATGCAACTGTTCAACAAACACCTCAGTCATTACACAGGCCTGTCCAAAGGTATCCTTTGAGCCAATTCCC
TTGTGGAGTCAGTAATGTGTCCGGACAGGTTTCCATAGGAACTCGGTTAAGGGATACATTATTGTGCCCCGGCTGGTTTTGCGATTCTAAAATGTAATAATAAGACG
TATGTAATAACACGGGGCCGACCAAAACGCTAAGATTTTACATTATTTCTGCTTCAATGGAACAGGACCATGTACAAATGTCAGCACAGTACAATGTACACATGGA
AAGTTACCTTGCTCTGGTACATGTTTACAGTCGTGTCATGTTACATGTGTACCTATTAGGCCAGTAGTATCAACTCAACTGCTGTAAATGGCAGTCTAGCAGAAGAA
TAATCCGGTCATCATAGTTGAGTTGACGACAATTTACCGTCAGATCGTCTTCTTGAGGTAGTAATTAGATCTGCCAATTTACAGACAATGCTAAAACCATTAATAGTA
CTCCATCATTAATCTAGACGGTTAAAGTGTCTGTTACGATTTTGGTATTATCAT

-58-

Table 11A (cont.)

CAGCTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAACAACAATACAAGA
GTCGACTTGGTTAGACATCTTTAATTAAACATGTTCTGGGTTGTTGTTATGTTCT

AAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTGTTACAATAGGAAAA
TTTTCATAGGCATAGGTCTCTCCTGGTCCCTCTCGTAAACAATGTTATCCTTTT

ATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCAAAATGGAATAAC
TATCCTTTATACTCTGTTTCGTGTAACATTGTAATCATCTCGTTTTACCTTATTG

ACTTTAAACAGATAGATAGCAAATTAAGAGAACAATTTGGAAATAATAAAACA
TGAAATTTTGTCTATCTATCGTTTAATTCTCTTGTTAAACCTTTATTATTTTGT

ATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTAACGCACAGTTTT
TATTAGAAATTCGTGAGGAGTCCTCCCTGGGTCTTTAACATTGCGTGTCAAAA

AATTGTGGAGGGGAATTTTTCTACTGTAATTCAACACAACCTGTTAATAGTACT
TTAACACCTCCCTTAAAAAGATGACATTAAGTTGTGTTGACAAATTATCATGA

TGGTTAATAGTACTTGGAGTACTAAAGGGTCAAATACACTGAAGGAAGTGAC
ACCAAATTATCATGAACCTCATGATTTCCAGTTTATTGTGACTTCCTTCACTG

ACAATCACCCCTCCCATGCAGAATAAAACAAATTATAAACATGTGGCAGGAAGTA
TGTTAGTGGGAGGGTACGTCTTATTTGTTTAATATTTGTACACCGTCCTTCAT

GGAAAAGCAATGTATGCCCCCTCCCATCAGTGGACAAATTAGATGTTTCATCAAT
CCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTTAATCTACAAGTAGTTTA

ATTACAGGGCTGCTATTAAACAAGAGATGGTGGTAATAGCAACAATGAGTCCGAG
TAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTGTTACTCAGGCTC

ATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGAAGTGAATTATAT
TAGAAGTCTGGACCTCCTCCTCTATACTCCCTGTTAACCTCTTCACTTAATATA

AAATATAAAGTAGTAAAAATTGAACCATTAGGAGTAGCACCCACCAAGGCAAAG
TTTATATTTTCATCATTTTTTAACCTTGGTAATCCTCATCGTGGGTGGTTCCGTTTC

AGAAGAGTGGTGCAGAGAGAAAAAGAGCAGTGGGAATAGGAGCTTTGTTCCCTT
TCTTCTCACACGTCTCTCTTTTTTCTCGTACCCCTTATCCTCGAAACAAGGAA

GGGTCTTGGGAGCAGCAGGAAGCACTATGGGCGCAGCGTCAATGACGCTGACG
CCCAAGAACCCTCGTCGTCTTTCGTGATACCCGCGTCGCAGTTACTGCGACTGC

GTACAGGCCAGACAATTATTGTCTGGTATAGTGCAGCAGCAGAACAAATTTGCTG
CATGTCCGGTCTGTTAATAACAGACCATATCACGTCGTCTGTGTTAAACGAC

AGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCACAGTCTGGGGCATCAAG
TCCCATAACTCCGCGTTGTCGTAGACAACGTTGAGTGTGAGACCCCGTAGTTC

CAGCTCCAGGCAAGAATCCTGGCTGTGGAAAGATACCTAAAGGATCAACAGCTC
GTCGAGGTCCGTTCTTAGGACCGACACCTTTCTATGGATTTCTAGTTGTGCGAG

-59-

Table 11A (cont.)

CTGGGGATTTGGGGTTGCTCTGGAAACTCATTTCACCACTGCTGTGCCTTGG
GACCCCTAAACCCAACGAGACCTTTTGAGTAAACGTGGTGACGACACGGAACC

AATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTTGGAATAACATGACCTGG
TTACGATCAACCTCATTATTTAGAGACCTTGTCTAAACCTTATTGTACTGGACC

ATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTCCCGGGAGCTCGAATT
TACCTCACCCCTGTCTCTTTAATTGTTAATGTGTTCTGAAGGGCCCTCGAGCTTAA

CTTGAAGACGAAAGGGCCTCG
GAACTTCTGCTTCCCGGAGC

-60-

CLAIMS

1. A recombinant DNA transfer vector comprising
DNA having all or part of the following nucleotide
sequence or equivalent nucleotide sequences containing
bases whose translated region codes for HTLV-III
envelope protein fragment denoted R10:
- ATGTTACGT
TACAATGCA
- CCTGTAGAAACCCCAACCCGTGAAATCAAAAACTCGACGGCCTGTGGGCATTCT
GGACATCTTTGGGGTTGGGCACCTTTAGTTTTTTGAGCTGCCGGACACCCGTAAG
- AGTCTGGATCGCGAAACTGTGGAATTGATCAATTCCCTGTGTGGAAGGAAGCA
TCAGACCTAGCGCTTTTGACACCTTAAGTTAAGGGACACACCTTCCTTCGT
- ACCACCACTCTATTTTGTGCATCAGATGCTAAAGCATATGATACAGAGGTACAT
TGGTGGTGAGATAAAACACGTAGTCTACGATTTCGTATACTATGTCTCCATGTA
- AATGTTTGGGCCACACATGCCTGTGTACCCACAGACCCCAACCCACAAGAAGTA
TTACAAACCCGGTGTGTACGGACACATGGGTGTCTGGGGTTGGGTGTTCTTCAT
- GTATTGGTAAATGTGACAGAAAATTTTAAACATGTGGAATAATGACATGGTAGAA
CATAACCATTACACTGTCTTTTAAATTGTACACCTTTTACTGTACCATCTT
- CAGATGCATGAGGATATAATCAGTTTATGGGATCAAAGCCTAAAGCCATGTGTA
GTCTACGTACTCCTATATTAGTCAAATACCTAGTTTCGGATTCGGTACACAT
- AAATTAACCCCACTCTGTGTTAGTTTAAAGTGCACTGATTTGAAGAATGATACT
TTTAATTGGGGTGAGACACAATCAAATTCACGTGACTAACTTCTTACTATGA
- AATACCAATAGTAGTAGCGGGAGAATGATAATGGAGAAAGGAGAGATAAAAAAC
TTATGGTTATCATCATCGCCCTCTTACTATTACCTCTTTCCTCTATTTTTTG
- TGCTCTTTCATATCAGCACAAGCATAAGAGGTAAGGTGCAGAAAGAATATGCA
ACGAGAAAGTTATAGTCGTGTTTCGTATTCTCCATTCCACGTCTTCTTATACGT
- TTTTTTTATAAACTTGATATAATACCAATAGATAATGATACTACCAGCTATACG
AAAAAATATTTGAACTATATTATGGTTATCTATTACTATGATGGTCGATATGC
- TTGACAAGTTGTAACACCTCAGTCATTACACAGGCCTGTCCAAAGGTATCCTTT
AACTGTTCAACATTGTGGAGTCAGTAATGTGTCCGGACAGGTTTCCATAGGAAA
- GAGCCAATTCCCATACATTATTGTGCCCGGCTGGTTTTTGCGATTCTAAAATGT
CTCGGTAAAGGTATGTAATAACACGGGGCCGACCAAACGCTAAGATTTTACA
- AATAATAAGACGTTCAATGGAACAGGACCATGTACAAATGTCAGCACAGTACAA
TTATTATTCTGCAAGTTACCTTGTCTGGTACATGTTTACAGTCGTGTCATGTT

-61-

34 TGTACACATGGAATTAGGCCAGTAGTATCAACTCAACTGCTGTTAAATGGCAGT
35 ACATGTGTACCTTAATCCGGTCATCATAGTTGAGTTGACGACAATTTACCGTCA
36 CTAGCAGAAGAAGAGGTAGTAATTAGATCTGCCAATTTACAGACAATGCTAAA
37 GATCGTCTTCTTCTCCATCATTAACTAGACGGTTAAAGTGTCTGTTACGATTT
38 ACCATAATAGTACAGCTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAAC
39 TGGTATTATCATGTGCGACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTTG
40 AACAAATACAAGAAAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTTGTT
41 TTGTTATGTTCTTTTTTCATAGGCATAGGTCTCTCCTAATCCCTCTCGTAAACAA
42 ACAATAGGAAAAATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCA
43 TGTTATCCTTTTTATCCTTTATACTCTGTTTCGTGTAAACATTGTAATCATCTCGT
44 AAATGGAATAACACTTTAAACAGATAGATAGCAAATTAAGAGAACAATTTGGA
45 TTACCTTATTGTGAAATTTTGTCTATCTATCGTTAATTCTCTTGTAAACCT
46 AATAATAAAACAATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTA
47 TTATTATTTTGTATTAGAAATTCGTCAGGAGTCCTCCCTGGGTCTTTAACAT
48 ACGCACAGTTTTAATTGTGGAGGGGAATTTTTCTACTGTAATTCAACACAACCTG
49 TGCGTGTCAAATTAACACCTCCCCTTAAAAAGATGACATTAAGTTGTGTTGAC
50 TTTAATAGTACTTGGTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACT
51 AAATTATCATGAACCAAATTATCATGAACCTCATGATTTCCCAGTTTATTGTGA
52 GAAGGAAGTGACACAATCACCTCCCATGCAGAATAAAACAAATTATAAACATG
53 CTTCTTCACTGTGTTAGTGGGAGGGTACGTCTTATTTTGTTTAATATTTGTAC
54 TGGCAGGAAGTAGGAAAAGCAATGTATGCCCCCTCCCATCAGTGGACAAATTAGA
55 ACCGTCCTTCATCCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTTAATCT
56 TGTTTCATCAAATATTACAGGGCTGCTATTAAACAAGAGATGGTGGTAATAGCAAC
57 ACAAGTAGTTTATAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTG
58 AATGAGTCCGAGATCCATCGCAGCGTAATGCTCTACACCACGCCGAACACCTGG
59 TTAATCAGGCTCTAGGTAGCGTCGCATTACGAGATGTGGTGCGGCTTGTGGACC
60 GTGGACGATATCACCGTGGTGACGCATGTGCGGCAAGACTGTAACCACGCGTCT
61 CACCTGCTATAGTGGCACCCTGCGTACAGCGCGTTCTGACATTGGTGCGCAGA
62 GTTGACTGGCAGGTGGTGGCCAATGGTGATGTCAGCGTTGAACTGCGTGATGCG
63 CAACTGACCGTCCACCACCGTTACCACTACAGTCGCAACTTGACGCACTACGC
64 GATCAACAGGTGGTTGCAACTGGACAAGGCACTAGCGGGACTTTGCAAGTGGTG
65 CTAGTTGTCCACCAACGTTGACCTGTTCCGTGATCGCCCTGAAACGTTCAACAC
66 AATCCGCACCTCTGGCAACCGGGTGAAGGTTATCTCTATGAACTGTGCGTCACA
67 TTAGGCGTGAGACCGTTGGCCCACTTCCAATAGAGATACTTGACACGCAGTGT
68 GCCAAAAGCCAGACAGAGTGTGATATCTACCCGCTTCGCGTCGGCATCCGGTCA
69 CGGTTTTTCGGTCTGTCTCACACTATAGATGGGCGAAGCGCAGCCGTAGGCCAGT

-62-

70 GTGGCAGTGAAGGGCGAACAGTTCCTGATTAAACCACAAACCGTTCTACTTTACT
71 CACCGTCACTTCCCGCTTGTC AAGGACTAATTGGTGT TTGGCAAGATGAAATGA
72 GGCTTTGGTCGT CATGAAGATGCGGACTTGCGTGGCAAAGGATTCGATAACGTG
73 CCGAAACCAGCAGCACTTCTACGCCTGAACGCACCGTTTTCTAAGCTATTGCAC
74 CTGATGGTGCACGACCACGCATTAATGGACTGGATTGGGGCCAACTCCTACCGT
75 GACTACCACGTGCTGGTGC GTAATTACCTGACCTAACCCCGGTTGAGGATGGCA
76 ACCTCGCATTACCCTTACGCTGAAGAGATGCTCGACTGGGCAGATGAACATGGC
77 TGGAGCGTAATGGGAATGCGACTTCTCTACGAGCTGACCCGCTCTACTTGTACCG
78 ATCGTGGTGATTGATGAAACTGCTGCTGTCGGCTTTAACCTCTCTTTAGGCATT
79 TAGCACCATAACTACTTTGACGACGACAGCCGAAATTGGAGAGAAATCCGTAA
80 GGTTTCGAAGCGGGCAACAAGCCGAAAGAACTGTACAGCGAAGAGGCAGTCAAC
81 CCAAAGCTTCGCCCCGTTGTTGCGCTTTCTTGACATGTCGCTTCTCCGT CAGTTG
82 GGGGAAACTCAGCAAGCGCACTTACAGGCGATTAAAGAGCTGATAGCGCGTGAC
83 CCCCTTTGAGTCGTTGCGGTGAATGTCCGCTAATTTCTCGACTATCGCGCACTG
84 AAAAACCACCCAAGCGTGGTGATGTGGAGTATTGCCAACGAACCGGATACCCGT
85 TTTTTGGTGGGTTGCAACCACTACACCTCATAACGGTTGCTTGGCCTATGGGCA
86 CCGCAAGGTGCACGGGAATATTTGCGGCCACTGGCGGAAGCAACGCGTAAACTC
87 GCGGTTCCACGTGCCCTTATAAAGCGCGGTGACCGCCTTCGTTGCGCATTTGAG
88 GACCCGACGCGTCCGATCACCTGCGTCAATGTAATGTTCTGCGACGCTCACACC
89 CTGGGCTGCGCAGGCTAGTGGACGCAGTTACATTACAAGACGCTGCGAGTGTGG
90 GATACCATCAGCGATCTCTTTGATGTGCTGTGCCTGAACCGTTATTACGGATGG
91 CTATGGTAGTCGCTAGAGAACTACACGACACGGACTTGGCAATAATGCCTACC
92 TATGTCCAAAGCGGCGATTTGGAAACGGCAGAGAAGGTACTGGAAAAAGAACTT
93 ATACAGGTTTCGCCGCTAAACCTTTGCCGTCTCTTCCATGACCTTTTTCTTGAA
94 CTGGCCTGGCAGGAGAAACTGCATCAGCCGATTATCATCACCGAATACGGCGTG
95 GACCGGACCGTCCTCTTTGACGTAGTCGGCTAATAGTAGTGGCTTATGCCGCAC
96 GATACGTTAGCCGGGCTGCACTCAATGTACACCGACATGTGGAGTGAAGAGTAT
97 CTATGCAATCGGCCCCGACGTGAGTTACATGTGGCTGTACACCTCACTTCTCATA
98 CAGTGTGCATGGCTGGATATGTATCACCGCGTCTTTGATCGCGTCAGCGCCGTC
99 GTCACACGTACCGACCTATACATAGTGGCGCAGAACTAGCGCAGTCGCGGCAG
100 GTCGGTGAACAGGTATGGAATTTGCGCCGATTTTGCGACCTCGCAAGGCATATTG
101 CAGCCACTTGTCATACCTTAAAGCGGCTAAACGCTGGAGCGTTCCGTATAAC
102 CGCGTTGGCGGTAAACAAGAAAGGGATCTTCACTCGCGACCGCAAACCGAAGTCG
103 GCGCAACCGCCATTGTTCTTTCCCTAGAAGTGAGCGCTGGCGTTTGGCTTCAGC
104 GCGGCTTTTCTGCTGCAAAAACGCTGGACTGGCATGAACTTCGGTGAAAAACCG
105 CGCCGAAAAGACGACGTTTTTTCGACCTGACCGTACTTGAAGCCACTTTTTGGC
106 CAGCAGGGAGGCAAACAA
107 GTCGTCCCTCCGTTTGTT.

1 2. A recombinant DNA transfer vector comprising
2 DNA having all or part of the following nucleotide
3 sequence or equivalent nucleotide sequences containing
4 bases whose translated region codes for HTLV-III
5 envelope protein fragment denoted PBI:

6 ATGTTACGTCCTGTAGAAACCCCAACCCGTGAAATCAAAAACTCGACGGCCTG
7 TACAATGCAGGACATCTTTGGGGTTGGGCACTTTAGTTTTTTGAGCTGCCGGAC
8 TGGGCATTTCAGTCTGGATCGCGAACCGGTGGCCGATCTGAACCAATCTGTAGAA
9 ACCCGTAAGTCAGACCTAGCGCTTGCGCACCGGCTAGACTTGGTTAGACATCTT
10 ATTAATTGTACAAGACCCAACAACAATACAAGAAAAAGTATCCGTATCCAGAGA
11 TAATTAACATGTTCTGGGTGTTGTTATGTTCTTTTTCATAGGCATAGGTCTCT
12 GGACCAGGGAGAGCATTGTTACAATAGGAAAAATAGGAAATATGAGACAAGCA
13 CCTAATCCCTCTCGTAAACAATGTTATCCTTTTTATCCTTTATACTCTGTTCTG
14 CATTGTAACATTAGTAGAGCAAAATGGAATAACACTTTAAAACAGATAGATAGC
15 GTAACATTGTAATCATCTCGTTTTACCTTATTGTGAAATTTTGTCTATCTATCG
16 AAATTAAGAGAACAATTTGGAAATAATAAAACAATAATCTTTAAGCAGTCCTCA
17 TTTAATTCTCTGTTAAACCTTTATTATTTTGTATTAGAAATTCGTCAGGAGT
18 GGAGGGGACCCAGAAATTGTAACGCACAGTTTTAATTGTGGAGGGGAATTTTTC
19 CCTCCCCCTGGGTCTTTAACATTGCGTGTCAAATTAACACCTCCCCTTAAAAAG
20 TACTGTAATTC AACACA ACTGTTAATAGTACTTG GTTTAATAGTACTTG GAGT
21 ATGACATTAAGTTGTGTTGACAAATTATCATGAACCAATTATCATGAACCTCA
22 ACTAAAGGGTCAAATAACACTGAAGGAAGTGACACAATCACCTCCCATGCAGA
23 TGATTTCCCAGTTTATTGTGACTTCCTTCACTGTGTTAGTGGGAGGGTACGTCT
24 ATAAAACAAATTATAAACATGTGGCAGGAAGTAGGAAAAGCAATGTATGCCCCCT
25 TATTTTGTTTAATATTTGTACACCGTCCTTCATCCTTTTCGTTACATACGGGGA
26 CCCATCAGTGGACAAATTAGATGTTTCATCAAATATTACAGGGCTGCTATTAAACA
27 GGGTAGTCACCTGTTAATCTACAAGTAGTTTATAATGTCCCGACGATAATTGT
28 AGAGATGGTGGTAATAGCAACAATGAGTCCGAGATCCGTCGACAAGCTTCCCGG
29 TCTCTACCACCATATCGTTGTTACTCAGGCTCTAGGCAGCTGTTTGAAGGGCC
30 GAGCTCGAATTCTTGAAGACGAAAGGGCCTCGTGATACTCCTATTTTTATAGGT
31 CTCGAGCTTAAGAACTTCTGCTTTCCCGGAGCACTATGCGGATAAAAATATCCA.

1 3. A recombinant DNA transfer vector comprising
2 DNA having all or part of the following nucleotide
3 sequence or equivalent nucleotide sequences containing
4 bases whose translated region codes for HTLV-III
5 envelope protein fragment denoted 590:

6 ATGTTACGTCCTGTAGAAAC
7 TACAATGCAGGACATCTTTGG

8 CCAACCCGTGAAATCAAAAACTCGACGGCCTGTGGGCATTCACTCTGGATCGC
9 GGTGGGCACTTTAGTTTTTTGAGATGCCGGACCCGTAAGTCAGACCTAGCG

10 GAACGCGTGGCCGATCTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAAC
11 CTTGCGCACCGGCTAGACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTTG

12 AACCAATACAAGAAAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTGT
13 TTGTTATGTTCTTTTTCATAGGCATAGGTCTCTCCTAATCCCTCTCGTAAACAA

14 ACAATAGGAAAAATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCA
15 TGTTATCCTTTTTATCCTTTATACTCTGTTTCGTGTAACATTGTAATCATCTCGT

16 AAATGGAATAACACTTTAAACAGATAGATAGCAAATTAAGAGAACAATTTGGA
17 TTTACCTTATTGTGAAATTTGTCTATCTATCGTTTAATTCTCTTGTTAAACCT

18 AATAATAAACAAATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTA
19 TTATTATTTTGTATTAGAAATTCGTCAGGAGTCCTCCCTGGGTCTTTAACAT

20 ACGCACAGTTTTAATTGTGGAGGGGAATTTTTCTACTGTAATTC AACACA ACTG
21 TGCGTGTCAAATTAACACCTCCCTTAAAAAGATGACATTAAGTTGTGTTGAC

22 TTTAATAGTACTTGGTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACT
23 AAATTATCATGAACCAAATTATCATGAACCTCATGATTTCCAGTTTATTGTGA

24 GAAGGAAGTGACACAATCACCTCCCATGCAGAATAAAACAAATTATAAACATG
25 CTTCTTCACTGTGTTAGTGGGAGGGTACGTCTATTTTGTTTAATATTTGTAC

26 TGGCAGGAAGTAGGAAAAGCAATGTATGCCCCCTCCCATCAGTGGACAAATTAGA
27 ACCGTCTTCATCCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTTAATCT

28 TGTTTCATCAAATATTACAGGGCTGCTATTAAACAAGAGATGGTGGTAATAGCAAC
29 ACAAGTAGTTTATAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTG

30 AATGAGTCCGAGATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGA
31 TTACTCAGGCTCTAGAAGTCTGGACCTCCTCCTCTATACTCCCTGTTAACCTCT

32 AGTGAATTATATAAATATAAAGTAGTAAAAATTGAACCATTAGGAGTAGCACCC
33 TCACTTAATATATTTATATTTTCATCATTTTTTAACCTTGGTAATCCTCATCGTGGG

34 ACCAAGGCAAAGAGAAGAGTGGTGCAGAGAGAAAAAAGAGCAGTGGGAATAGGA
35 TGGTTCCGTTTCTCTTCTCACCACGTCTCTCTTTTTTCTCGTCACCTTATCCT

-65-

36 GCTTTGTTCTTGGGTTCTTGGGAGCAGCAGGAAGCACTATGGGCGCAGCGTCA
37 CGAAACAAGGAACCCAAGAACCCTCGTCGTCTTCGTGATACCCGCGTCGCAGT
38 ATGACGCTGACGGTACAGGCCAGACAATTATTGTCTGGTATAGTGCAGCAGCAG
39 TACTGCGACTGCCATGTCCGGTCTGTTAATAACAGACCATATCACGTCGTCGTC
40 AACAAATTGCTGAGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCACAGTC
41 TTGTTAAACGACTCCCGATAACTCCGCGTTGTCGTAGACAACGTTGAGTGTCAG
42 TGGGGCATCAAGCAGCTCCAGGCAAGAATCCTGGCTGTGGAAGATACCTAAAG
43 ACCCCGTAGTTCGTGAGGTCCGTTCTTAGGACCGACACCTTTCTATGGATTTC
44 GATCAACAGCTCCTGGGGATTTGGGGTTGCTCTGGAAACTCATTTGCACCACT
45 CTAGTTGTCGAGGACCCCTAAACCCCAACGAGACCTTTTGAGTAAACGTGGTGA
46 GCTGTGCCTTGAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTTGGAAT
47 CGACACGGAACCTTACGATCAACCTCATTATTTAGAGACCTTGTCTAAACCTTA
48 AACATGACCTGGATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTCCCG
49 TTGTACTGGACCTACCTCACCCCTGTCTCTTTAATTGTTAATGTGTTGGAAGGGC
50 ATCCATCGCAGCGTAATGCTCTACACCACGCCGAACACCTGGGTGGACGATATC
51 TAGGTAGCGTCGCATTACGAGATGTGGTGCGGCTTGTGGACCCACCTGCTATAG
52 ACCGTGGTGACGCATGTGCGCAAGACTGTAACCACGCGTCTGTTGACTGGCAG
53 TGGCACCACCTGCGTACAGCGCGTTCTGACATTGGTGCGCAGACAACCTGACCGTC
54 GTGGTGGCCAATGGTGATGTCAGCGTTGAACTGCGTGATGCGGATCAACAGGTG
55 CACCACCGGTTACCACTACAGTCGCAACTTGACGCACTACGCCTAGTTGTCCAC
56 GTTGCAACTGGACAAGGCACTAGCGGGACTTTGCAAGTGGTGAATCCGCACCTC
57 CAACGTTGACCTGTTCCGTGATCGCCCTGAAACGTTACCACTTAGGCGTGGAG
58 TGGCAACCGGGTGAAGGTTATCTCTATGAACTGTGCGTCACAGCCAAAAGCCAG
59 ACCGTTGGCCCACTTCCAATAGAGATACTTGACACGCAGTGTGCGTTTTGCGTC
60 ACAGAGTGTGATATCTACCCGCTTCGCGTCGGCATCCGGTCAGTGGCAGTGAAG
61 TGTCTCACACTATAGATGGGCGAAGCGCAGCCGTAGGCCAGTCACCGTCACTTC
62 GGCGAACAGTTCCTGATTAACCACAAACCGTTCTACTTTACTGGCTTTGGTCGT
63 CCGCTTGTCAAGGACTAATTGGTGTTTGGCAAGATGAAATGACCGAAACCAGCA
64 CATGAAGATGCGGACTTGCGTGGCAAAGGATTGCGATAACGTGCTGATGGTGCAC
65 GCACTTCTACGCCTGAACGCACCGTTTCCTAAGCTATTGCACGACTACCACGTG
66 GACCACGCATTAATGGACTGGATTGGGGCCAACTCCTACCGTACCTCGCATTAC
67 CTGGTGCGTAATTACCTGACCTAACCCCGTTGAGGATGGCATGGAGCGTAATG
68 CCTTACGCTGAAGAGATGCTCGACTGGGCAGATGAACATGGCATCGTGGTGATT
69 GGAATGCGACTTCTCTACGAGCTGACCCGTCTACTTGTACCGTAGCACCCTAA
70 GATGAACTGCTGCTGTGCGGCTTTAACCTCTCTTTAGGCATTGGTTTCGAAGCG
71 CTACTTTGACGACGACAGCCGAAATTGGAGAGAAATCCGTAACCAAAGCTTCGC

-66-

72 GGCAACAAGCCGAAAGAAGTGTACAGCGAAGAGGCAGTCAACGGGGAAACTCAG
 73 CCGTTGTTGCGCTTTCTTGACATGTCGCTTCTCCGTCAGTTGCCCTTTGAGTC

 74 CAAGCGCACTTACAGGCGATTAAAGAGCTGATAGCGCGTGACAAAAACCACCCA
 75 GTTCGCGTGAATGTCCGCTAATTTCTCGACTATCGCGCACTGTTTTTGGTGGGT

 76 AGCGTGGTGATGTGGAGTATTGCCAACGAACCGGATACCCGTCGCAAGGTGCA
 77 TCGCACCCTACACCTCATAACGGTTGCTTGGCCTATGGGCAGGCGTTCCACGT

 78 CGGGAATATTTGCGGCCACTGGCGGAAGCAACGCGTAAACTCGACCCGACGCGT
 79 GCCCTTATAAAGCGCGGTGACCGCCTTCGTTGCGCATTTGAGCTGGGCTGCGCA

 80 CCGATCACCTGCGTCAATGTAATGTTCTGCGACGCTCACACCGATACCATCAGC
 81 GGCTAGTGGACGCAGTTACATTACAAGACGCTGCGAGTGTGGCTATGGTAGTCG

 82 GATCTCTTTGATGTGCTGTGCCTGAACCGTTATTACGGATGGTATGTCCAAAGC
 83 CTAGAGAACTACACGACACGGACTTGGCAATAATGCCTACCATACAGGTTTCG

 84 GGCGATTTGGAAACGGCAGAGAAGGTACTGGAAAAAGAACTTCTGGCCTGGCAG
 85 CCGCTAAACCTTTGCGCTCTCTCCATGACCTTTTTCTTGAAGACCGGACCGTC

 86 GAGAACTGCATCAGCCGATTATCATCACCGAATACGGCGTGGATACGTTAGCC
 87 CTCTTTGACGTAGTCGGCTAATAGTAGTGGCTTATGCCGCACCTATGCAATCGG

 88 GGGCTGCACTCAATGTACACCGACATGTGGAGTGAAGAGTATCAGTGTGCATGG
 89 CCCGACGTGAGTTACATGTGGCTGTACACCTCACTTCTCATAGTCACACGTACC

 90 CTGGATATGTATCACCGCGTCTTTGATCGCGTCAGCGCCGTCGTCGGTGAACAG
 91 GACCTATACATAGTGGCGCAGAACTAGCGCAGTCGCGGCAGCAGCCACTTGTC

 92 GTATGGAATTTGCGCGATTTTGCGACCTCGCAAGGCATATTGCGCGTTGGCGGT
 93 CATACTTAAAGCGGCTAAACGCTGGAGCGTTCGCTATAACGCGCAACCGCCA

 94 AACAAGAAAGGGATCTTCACTCGCGACCGCAAACCGAAGTCGGCGGCTTTTCTG
 95 TTGTTCTTTCCCTAGAAGTGAGCGCTGGCGTTTGGCTTCAGCCGCCGAAAAGAC

 96 CTGCAAAAACGCTGGACTGGCATGAACTTCGGTGAAAAACCGCAGCAGGGAGGC
 97 GACGTTTTTGCGACCTGACCGTACTTGAAGCCACTTTTTGGCGTCGTCCCTCCG

 98 AAACAA
 99 TTTGTT.

- 1 4. A recombinant DNA transfer vector comprising
- 2 DNA having all or part of the following nucleotide
- 3 sequence or equivalent nucleotide sequences containing
- 4 bases whose translated region codes for HTLV-III
- 5 envelope protein fragment denoted KHI:

-67-

6 ATGTTACGT
7 TACAATGCA
8 CCTGTAGAAACCCCAACCCGTGAAATCAAAAACTCGACGGCCTGTGGGCATTC
9 GGACATCTTTGGGGTTGGGCACTTTAGTTTTTTGAGCTGCCGGACACCCGTAAG
10 AGTCTGGATCGCGAACGCGAATTCCTGTGTGGAAGGAAGCAACCACCACTCTA
11 TCAGACCTAGCGCTTGCGCTTAAGGGACACACCTTCCTTCGTTGGTGGTGAGAT
12 TTTTGTGCATCAGATGCTAAAGCATATGATACAGAGGTACATAATGTTTGGGCC
13 AAAACACGTAGTCTACGATTTTCGTATACTATGTCTCCATGTATTACAAACCCGG
14 ACACATGCCTGTGTACCCACAGACCCCAACCCACAAGAAGTAGTATTGGTAAAT
15 TGTGTACGGACACATGGGTGTCTGGGGTTGGGTGTTCTTCATCATAACCATTTA
16 GTGACAGAAAATTTTAAACATGTGGAAAAATGACATGGTAGAACAGATGCATGAG
17 CACTGTCTTTTAAATTTGTACACCTTTTACTGTACCATCTTGTCTACGTACTC
18 GATATAATCAGTTTATGGGATCAAAGCCTAAAGCCATGTGTAAAATTAACCCCA
19 CTATATTAGTCAAATACCCTAGTTTCGGATTTCCGTACACATTTTAATTGGGGT
20 CTCTGTGTTAGTTTAAAGTGCCTGATTTGAAGAATGATACTAATACCAATAGT
21 GAGACACAATCAAATTTACGTGACTAACTTCTTACTATGATTATGGTTATCA
22 AGTAGCGGGAGAATGATAATGGAGAAAGGAGAGATAAAAACTGCTCTTTCAAT
23 TCATCGCCCTCTTACTATTACCTCTTCCCTCTCTATTTTTTTGACGAGAAAGTTA
24 ATCAGCACAAGCATAAGAGGTAAGGTGCAGAAAGAATATGCATTTTTTTTATAAA
25 TAGTCGTGTTTCGTATTCTCCATTCCACGTCTTCTTATACGTAAAAAATATTT
26 CTTGATATAATACCAATAGATAATGATACTACCAGCTATACGTTGACAAGTTGT
27 GAACTATATTATGGTTATCTATTACTATGATGGTCGATATGCAACTGTTCAACA
28 AACACCTCAGTCATTACACAGGCCTGTCCAAAGGTATCCTTTGAGCCAATTCCC
29 TTGTGGAGTCAGTAATGTGTCCGGACAGGTTTCCATAGGAACTCGGTTAAGGG
30 ATACATTATTGTGCCCCGGCTGGTTTTGCGATTCTAAAATGTAATAATAAGACG
31 TATGTAATAACACGGGGCCGACCAAAACGCTAAGATTTTACATTATTATTCTGC
32 TTCAATGGAACAGGACCATGTACAAATGTCAGCACAGTACAATGTACACATGGA
33 AAGTTACCTTGTCCTGGTACATGTTTACAGTCGTGTCATGTTACATGTGTACCT
34 ATTAGGCCAGTAGTATCAACTCAACTGCTGTAAATGGCAGTCTAGCAGAAGAA
35 TAATCCGGTCATCATAGTTGAGTTGACGACAATTTACCGTCAGATCGTCTTCTT
36 GAGGTAGTAATTAGATCTGCCAATTTACAGACAATGCTAAAACCATAATAGTA
37 CTCCATCATTAATCTAGACGGTTAAAGTGTCTGTTACGATTTTGGTATTATCAT
38 CAGCTGAACCAATCTGTAGAAATTAATTGTACAAGACCCAACAATAACAAGA
39 GTCGACTTGGTTAGACATCTTTAATTAACATGTTCTGGGTGTTGTTATGTTCT
40 AAAAGTATCCGTATCCAGAGAGGACCAGGGAGAGCATTTGTTACAATAGGAAAA
41 TTTTCATAGGCATAGGTCTCTCCTGGTCCCTCTCGTAAACAATGTTATCCTTTT

-68-

42 ATAGGAAATATGAGACAAGCACATTGTAACATTAGTAGAGCAAAATGGAATAAC
43 TATCCTTTATACTCTGTTCGTGTAACATTGTAATCATCTCGTTTTACCTTATTG
44 ACTTTAAACAGATAGATAGCAAATTAAGAGAACAAATTTGGAAATAATAAAACA
45 TGAAATTTTGTCTATCTATCGTTTAATTCTCTTGTTAAACCTTTATTATTTTGT
46 ATAATCTTTAAGCAGTCCTCAGGAGGGGACCCAGAAATTGTAACGCACAGTTTT
47 TATTAGAAATTCGT CAGGAGTCCTCCCTGGGTCTTAAACATTGCGTGTCAAAA
48 AATTGTGGAGGGGAATTTTCTACTGTAATTCAACACAACCTGTTTAATAGTACT
49 TTAACACCTCCCTTAAAAAGATGACATTAAGTTGTGTTGACAAATTATCATGA
50 TGGTTTAATAGTACTTGGAGTACTAAAGGGTCAAATAACACTGAAGGAAGTGAC
51 ACCAAATTATCATGAACCTCATGATTCCCAGTTTATTGTGACTTCCTTCACTG
52 ACAATCACCTCCCATGCAGAATAAAACAAATTATAAACATGTGGCAGGAAGTA
53 TGTTAGTGGGAGGGTACGTCTTATTTGTTTAATATTTGTACACCGTCCTTCAT
54 GGAAAAGCAATGTATGCCCTCCCATCAGTGGACAAATTAGATGTTTCATCAAA
55 CCTTTTCGTTACATACGGGGAGGGTAGTCACCTGTTTAATCTACAAGTAGTTTA
56 ATTACAGGGCTGCTATTAACAAGAGATGGTGGTAATAGCAACAATGAGTCCGAG
57 TAATGTCCCGACGATAATTGTTCTCTACCACCATTATCGTTGTTACTCAGGCTC
58 ATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGAAAGTGAATTATAT
59 TAGAAGTCTGGACCTCCTCCTCTATACTCCCTGTTAACCTCTTCACTTAATATA
60 AAATATAAAGTAGTAAAAATTGAACCATTAGGAGTAGCACCCACCAAGGCAAAG
61 TTTATATTTTCATCATTTTTAACTTGTAATCCTCATCGTGGGTGGTTCCGTTTC
62 AGAAGAGTGGTGCAGAGAGAAAAAGAGCAGTGGGAATAGGAGCTTTGTTCCCTT
63 TCTTCTCACCACGTCTCTTTTTTCTCGTCACCCTTATCCTCGAAACAAGGAA
64 GGGTTCTTGGGAGCAGCAGGAAGCACTATGGGCGCAGCGTCAATGACGCTGACG
65 CCAAGAACCCTCGTCGTCTTCGTGATACCCGCGTCGCAGTTACTGCGACTGC
66 GTACAGGCCAGACAATTATTGTCTGGTATAGTGCAGCAGCAGAACAAATTTGCTG
67 CATGTCCGGTCTGTTAATAACAGACCATATCACGTCGTCTGTTAAACGAC
68 AGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCACAGTCTGGGGCATCAAG
69 TCCCGATAACTCCGCGTTGTCTGAGACAACGTTGAGTGTGAGACCCCGTAGTTC
70 CAGCTCCAGGCAAGAATCCTGGCTGTGGAAAGATACCTAAAGGATCAACAGCTC
71 GTCGAGGTCCGTTCTTAGGACCGACACCTTTCTATGGATTTCTAGTTGTGCGAG
72 CTGGGGATTTGGGGTTGCTCTGGAAACTCATTTGCACCACTGCTGTGCCTTGG
73 GACCCCTAAACCCCAACGAGACCTTTGAGTAAACGTGGTGACGACACGGAACC
74 AATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTTGGAATAACATGACCTGG
75 TTACGATCAACCTCATTTATTAGAGACCTTGCTAAACCTTATTGTACTGGACC
76 ATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTCCCGGGAGCTCGAATT
77 TACCTCACCTGTCTCTTTAATTGTTAATGTGTTGGAAGGGCCCTCGAGCTTAA
78 CTTGAAGACGAAAGGGCCTCG
79 GAACTTCTGCTTTCCCGGAGC.

5. The DNA transfer vector of any preceding claim transferred to and replicated in a eukaryotic or prokaryotic host.

6. A host transformed by the transfer vector of any of claims 1 to 4.

7. HTLV-III envelope protein fragment denoted R10 having the following amino-acid sequence, or mutants thereof:

MetLeuArg

10 ProValGluThrProThrArgGluIleLysLysLeuAspGlyLeuTrpAlaPhe
SerLeuAspArgGluAsnCysGlyIleAspGlnPheProValTrpLysGluAla
ThrThrThrLeuPheCysAlaSerAspAlaLysAlaTyrAspThrGluValHis
AsnValTrpAlaThrHisAlaCysValProThrAspProAsnProGlnGluVal
15 ValLeuValAsnValThrGluAsnPheAsnMetTrpLysAsnAspMetValGlu

20

25

30

35

-70-

10 GlnMetHisGluAspIleIleSerLeuTrpAspGlnSerLeuLysProCysVal
11 LysLeuThrProLeuCysValSerLeuLysCysThrAspLeuLysAsnAspThr
12 AsnThrAsnSerSerSerGlyArgMetIleMetGluLysGlyGluIleLysAsn
13 CysSerPheAsnIleSerThrSerIleArgGlyLysValGlnLysGluTyrAla
14 PhePheTyrLysLeuAspIleIleProIleAspAsnAspThrThrSerTyrThr
15 LeuThrSerCysAsnThrSerValIleThrGlnAlaCysProLysValSerPhe
16 GluProIleProIleHisTyrCysAlaProAlaGlyPheAlaIleLeuLysCys
17 AsnAsnLysThrPheAsnGlyThrGlyProCysThrAsnValSerThrValGln
18 CysThrHisGlyIleArgProValValSerThrGlnLeuLeuLeuAsnGlySer
19 LeuAlaGluGluGluValValIleArgSerAlaAsnPheThrAspAsnAlaLys
20 ThrIleIleValGlnLeuAsnGlnSerValGluIleAsnCysThrArgProAsn
21 AsnAsnThrArgLysSerIleArgIleGlnArgGlyProGlyArgAlaPheVal
22 ThrIleGlyLysIleGlyAsnMetArgGlnAlaHisCysAsnIleSerArgAla
23 LysTrpAsnAsnThrLeuLysGlnIleAspSerLysLeuArgGluGlnPheGly
24 AsnAsnLysThrIleIlePheLysGlnSerSerGlyGlyAspProGluIleVal
25 ThrHisSerPheAsnCysGlyGlyGluPhePheTyrCysAsnSerThrGlnLeu
26 PheAsnSerThrTrpPheAsnSerThrTrpSerThrLysGlySerAsnAsnThr
27 GluGlySerAspThrIleThrLeuProCysArgIleLysGlnIleIleAsnMet
28 TrpGlnGluValGlyLysAlaMetTyrAlaProProIleSerGlyGlnIleArg
29 CysSerSerAsnIleThrGlyLeuLeuLeuThrArgAspGlyGlyAsnSerAsn
30 AsnGluSerGluIleHisArgSerValMetLeuTyrThrThrProAsnThrTrp
31 ValAspAspIleThrValValThrHisValAlaGlnAspCysAsnHisAlaSer
32 ValAspTrpGlnValValAlaAsnGlyAspValSerValGluLeuArgAspAla
33 AspGlnGlnValValAlaThrGlyGlnGlyThrSerGlyThrLeuGlnValVal
34 AsnProHisLeuTrpGlnProGlyGluGlyTyrLeuTyrGluLeuCysValThr
35 AlaLysSerGlnThrGluCysAspIleTyrProLeuArgValGlyIleArgSer

-71-

36 ValAlaValLysGlyGluGlnPheLeuIleAsnHisLysProPheTyrPheThr
 37 GlyPheGlyArgHisGluAspAlaAspLeuArgGlyLysGlyPheAspAsnVal
 38 LeuMetValHisAspHisAlaLeuMetAspTrpIleGlyAlaAsnSerTyrArg
 39 ThrSerHisTyrProTyrAlaGluGluMetLeuAspTrpAlaAspGluHisGly
 40 IleValValIleAspGluThrAlaAlaValGlyPheAsnLeuSerLeuGlyIle
 41 GlyPheGluAlaGlyAsnLysProLysGluLeuTyrSerGluGluAlaValAsn
 42 GlyGluThrGlnGlnAlaHisLeuGlnAlaIleLysGluLeuIleAlaArgAsp
 43 LysAsnHisProSerValValMetTrpSerIleAlaAsnGluProAspThrArg
 44 ProGlnGlyAlaArgGluTyrPheAlaProLeuAlaGluAlaThrArgLysLeu
 45 AspProThrArgProIleThrCysValAsnValMetPheCysAspAlaHisThr
 46 AspThrIleSerAspLeuPheAspValLeuCysLeuAsnArgTyrTyrGlyTrp
 47 TyrValGlnSerGlyAspLeuGluThrAlaGluLysValLeuGluLysGluLeu
 48 LeuAlaTrpGlnGluLysLeuHisGlnProIleIleIleThrGluTyrGlyVal
 49 AspThrLeuAlaGlyLeuHisSerMetTyrThrAspMetTrpSerGluGluTyr
 50 GlnCysAlaTrpLeuAspMetTyrHisArgValPheAspArgValSerAlaVal
 51 ValGlyGluGlnValTrpAsnPheAlaAspPheAlaThrSerGlnGlyIleLeu
 52 ArgValGlyGlyAsnLysLysGlyIlePheThrArgAspArgLysProLysSer
 53 AlaAlaPheLeuLeuGlnLysArgTrpThrGlyMetAsnPheGlyGluLysPro
 54 GlnGlnGlyGlyLysGln.

1 8. HTLV-III envelope protein fragment denoted
 2 PBl having the following amino-acid sequence, or
 3 mutants thereof:
 4 MetLeuArg
 5 ProValGluThrProThrArgGluIleLysLysLeuAspGlyLeuTrpAlaPhe
 6 SerLeuAspArgGluArgValAlaAspLeuAsnGlnSerValGluIleAsnCys
 7 ThrArgProAsnAsnAsnThrArgLysSerIleArgIleGlnArgGlyProGly

-72-

8 ArgAlaPheValThrIleGlyLysIleGlyAsnMetArgGlnAlaHisCysAsn
 9 IleSerArgAlaLysTrpAsnAsnThrLeuLysGlnIleAspSerLysLeuArg
 10 GluGlnPheGlyAsnAsnLysThrIleIlePheLysGlnSerSerGlyGlyAsp
 11 ProGluIleValThrHisSerPheAsnCysGlyGlyGluPhePheTyrCysAsn
 12 SerThrGlnLeuPheAsnSerThrTrpPheAsnSerThrTrpSerThrLysGly
 13 SerAsnAsnThrGluGlySerAspThrIleThrLeuProCysArgIleLysGln
 14 IleIleAsnMetTrpGlnGluValGlyLysAlaMetTyrAlaProProIleSer
 15 GlyGlnIleArgCysSerSerAsnIleThrGlyLeuLeuLeuThrArgAspGly
 16 GlyAsnSerAsnAsnGluSerGluIleArgArgGlnAlaSerArgGluLeuGlu
 17 PheLeuLysThrLysGlyProArgAspThrProIlePheIleGly.

1 9. HTLV-III envelope protein fragment denoted 590
 2 having the following amino-acid sequence, or mutants
 3 thereof:

4 MetLeuArgProValGluThr
 5 ProThrArgGluIleLysLysLeuAspGlyLeuTrpAlaPheSerLeuAspArg
 6 GluArgValAlaAspLeuAsnGlnSerValGluIleAsnCysThrArgProAsn
 7 AsnAsnThrArgLysSerIleArgIleGlnArgGlyProGlyArgAlaPheVal
 8 ThrIleGlyLysIleGlyAsnMetArgGlnAlaHisCysAsnIleSerArgAla
 9 LysTrpAsnAsnThrLeuLysGlnIleAspSerLysLeuArgGluGlnPheGly
 10 AsnAsnLysThrIleIlePheLysGlnSerSerGlyGlyAspProGluIleVal
 11 ThrHisSerPheAsnCysGlyGlyGluPhePheTyrCysAsnSerThrGlnLeu
 12 PheAsnSerThrTrpPheAsnSerThrTrpSerThrLysGlySerAsnAsnThr
 13 GluGlySerAspThrIleThrLeuProCysArgIleLysGlnIleIleAsnMet
 14 TrpGlnGluValGlyLysAlaMetTyrAlaProProIleSerGlyGlnIleArg
 15 CysSerSerAsnIleThrGlyLeuLeuLeuThrArgAspGlyGlyAsnSerAsn
 16 AsnGluSerGluIlePheArgProGlyGlyGlyAspMetArgAspAsnTrpArg
 17 SerGluLeuTyrLysTyrLysValValLysIleGluProLeuGlyValAlaPro

18 ThrLysAlaLysArgArgValValGlnArgGluLysArgAlaValGlyIleGly
19 AlaLeuPheLeuGlyPheLeuGlyAlaAlaGlySerThrMetGlyAlaAlaSer
20 MetThrLeuThrValGlnAlaArgGlnLeuLeuSerGlyIleValGlnGlnGln
21 AsnAsnLeuLeuArgAlaIleGluAlaGlnGlnHisLeuLeuGlnLeuThrVal
22 TrpGlyIleLysGlnLeuGlnAlaArgIleLeuAlaValGluArgTyrLeuLys
23 AspGlnGlnLeuLeuGlyIleTrpGlyCysSerGlyLysLeuIleCysThrThr
24 AlaValProTrpAsnAlaSerTrpSerAsnLysSerLeuGluGlnIleTrpAsn
25 AsnMetThrTrpMetGluTrpAspArgGluIleAsnAsnTyrThrSerPhePro
26 IleHisArgSerValMetLeuTyrThrThrProAsnThrTrpValAspAspIle
27 ThrValValThrHisValAlaGlnAspCysAsnHisAlaSerValAspTrpGln
28 ValValAlaAsnGlyAspValSerValGluLeuArgAspAlaAspGlnGlnVal
29 ValAlaThrGlyGlnGlyThrSerGlyThrLeuGlnValValAsnProHisLeu
30 TrpGlnProGlyGluGlyTyrLeuTyrGluLeuCysValThrAlaLysSerGln
31 ThrGluCysAspIleTyrProLeuArgValGlyIleArgSerValAlaValLys
32 GlyGluGlnPheLeuIleAsnHisLysProPheTyrPheThrGlyPheGlyArg
33 HisGluAspAlaAspLeuArgGlyLysGlyPheAspAsnValLeuMetValHis
34 AspHisAlaLeuMetAspTrpIleGlyAlaAsnSerTyrArgThrSerHisTyr
35 ProTyrAlaGluGluMetLeuAspTrpAlaAspGluHisGlyIleValValIle
36 AspGluThrAlaAlaValGlyPheAsnLeuSerLeuGlyIleGlyPheGluAla
37 GlyAsnLysProLysGluLeuTyrSerGluGluAlaValAsnGlyGluThrGln
38 GlnAlaHisLeuGlnAlaIleLysGluLeuIleAlaArgAspLysAsnHisPro
39 SerValValMetTrpSerIleAlaAsnGluProAspThrArgProGlnGlyAla
40 ArgGluTyrPheAlaProLeuAlaGluAlaThrArgLysLeuAspProThrArg
41 ProIleThrCysValAsnValMetPheCysAspAlaHisThrAspThrIleSer
42 AspLeuPheAspValLeuCysLeuAsnArgTyrTyrGlyTrpTyrValGlnSer
43 GlyAspLeuGluThrAlaGluLysValLeuGluLysGluLeuLeuAlaTrpGln
44 GluLysLeuHisGlnProIleIleIleThrGluTyrGlyValAspThrLeuAla

-74-

45 GlyLeuHisSerMetTyrThrAspMetTrpSerGluGluTyrGlnCysAlaTrp
 46 LeuAspMetTyrHisArgValPheAspArgValSerAlaValValGlyGluGln
 47 ValTrpAsnPheAlaAspPheAlaThrSerGlnGlyIleLeuArgValGlyGly
 48 AsnLysLysGlyIlePheThrArgAspArgLysProLysSerAlaAlaPheLeu
 49 LeuGlnLysArgTrpThrGlyMetAsnPheGlyGluLysProGlnGlnGlyGly
 50 LysGln.

1 10. HTLV-III envelope protein fragment denoted
 2 KHI having the following amino-acid sequence, or
 3 mutants thereof:

4 MetLeuArg
 5 ProValGluThrProThrArgGluIleLysLysLeuAspGlyLeuTrpAlaPhe
 6 SerLeuAspArgGluArgGluPheProValTrpLysGluAlaThrThrThrLeu
 7 PheCysAlaSerAspAlaLysAlaTyrAspThrGluValHisAsnValTrpAla
 8 ThrHisAlaCysValProThrAspProAsnProGlnGluValValLeuValAsn
 9 ValThrGluAsnPheAsnMetTrpLysAsnAspMetValGluGlnMetHisGlu
 10 AspIleIleSerLeuTrpAspGlnSerLeuLysProCysValLysLeuThrPro
 11 LeuCysValSerLeuLysCysThrAspLeuLysAsnAspThrAsnThrAsnSer
 12 SerSerGlyArgMetIleMetGluLysGlyGluIleLysAsnCysSerPheAsn
 13 IleSerThrSerIleArgGlyLysValGlnLysGluTyrAlaPhePheTyrLys
 14 LeuAspIleIleProIleAspAsnAspThrThrSerTyrThrLeuThrSerCys
 15 AsnThrSerValIleThrGlnAlaCysProLysValSerPheGluProIlePro
 16 IleHisTyrCysAlaProAlaGlyPheAlaIleLeuLysCysAsnAsnLysThr
 17 PheAsnGlyThrGlyProCysThrAsnValSerThrValGlnCysThrHisGly
 18 IleArgProValValSerThrGlnLeuLeuLeuAsnGlySerLeuAlaGluGlu
 19 GluValValIleArgSerAlaAsnPheThrAspAsnAlaLysThrIleIleVal
 20 GlnLeuAsnGlnSerValGluIleAsnCysThrArgProAsnAsnAsnThrArg
 21 LysSerIleArgIleGlnArgGlyProGlyArgAlaPheValThrIleGlyLys

-75-

22 IleGlyAsnMetArgGlnAlaHisCysAsnIleSerArgAlaLysTrpAsnAsn
 23 ThrLeuLysGlnIleAspSerLysLeuArgGluGlnPheGlyAsnAsnLysThr
 24 IleIlePheLysGlnSerSerGlyGlyAspProGluIleValThrHisSerPhe
 25 AsnCysGlyGlyGluPhePheTyrCysAsnSerThrGlnLeuPheAsnSerThr
 26 TrpPheAsnSerThrTrpSerThrLysGlySerAsnAsnThrGluGlySerAsp
 27 ThrIleThrLeuProCysArgIleLysGlnIleIleAsnMetTrpGlnGluVal
 28 GlyLysAlaMetTyrAlaProProIleSerGlyGlnIleArgCysSerSerAsn
 29 IleThrGlyLeuLeuLeuThrArgAspGlyGlyAsnSerAsnAsnGluSerGlu
 30 IlePheArgProGlyGlyGlyAspMetArgAspAsnTrpArgSerGluLeuTyr
 31 LysTyrLysValValLysIleGluProLeuGlyValAlaProThrLysAlaLys
 32 ArgArgValValGlnArgGluLysArgAlaValGlyIleGlyAlaLeuPheLeu
 33 GlyPheLeuGlyAlaAlaGlySerThrMetGlyAlaAlaSerMetThrLeuThr
 34 ValGlnAlaArgGlnLeuLeuSerGlyIleValGlnGlnGlnAsnAsnLeuLeu
 35 ArgAlaIleGluAlaGlnGlnHisLeuLeuGlnLeuThrValTrpGlyIleLys
 36 GlnLeuGlnAlaArgIleLeuAlaValGluArgTyrLeuLysAspGlnGlnLeu
 37 LeuGlyIleTrpGlyCysSerGlyLysLeuIleCysThrThrAlaValProTrp
 38 AsnAlaSerTrpSerAsnLysSerLeuGluGlnIleTrpAsnAsnMetThrTrp
 39 MetGluTrpAspArgGluIleAsnAsnTyrThrSerPheProGlyAlaArgIle
 40 LeuGluAspGluArgAlaSer.

- 1 11. A plasmid selected from the following:
- 2 plasmid pREV1, plasmid pREV1TT, plasmid pREV1TT/ch1,
- 3 plasmid pREV2.2, plasmid pR10, plasmid pPB1, plasmid
- 4 p590, and plasmid pKH1,

and preferably any of the last five of these eight plasmids.

12. DNA having the nucleotide sequence defined in any of claims 1 to 4, or an equivalent nucleotide sequence
5 containing bases whose translated region codes for HTLV-III envelope protein fragment denoted R10, PB1, 590 or KH1.

13. An immunochemical assay for detecting or quantifying antibody against HTLV-III in a fluid, which comprises
10 employing an HTLV-III protein selected from R10, PB1, 590 and KH1.

14. An immunoabsorbent suitable for use in a solid phase immunochemical assay for antibody against HTLV-III, which comprises a solid phase to which is affixed an HTLV-III
15 protein selected from R10, PB1, 590 and KH1.

15. An immunoabsorbent according to claim 14, wherein the solid phase is a glass or plastic bead, a well of a microtiter plate or a test tube.

16. An immunoabsorbent according to claim 14 or claim
20 15, which additionally comprises a post-coat of animal protein.

17. A kit suitable for use in detecting antibody against HTLV-III in a biological fluid, which comprises:

(a) an immunoabsorbent according to any of claims
25 14 to 16;

(b) labelled HTLV-III antibody; and

(c) means for detecting the label associated with the immunoabsorbent.

18. A kit according to claim 17, wherein the
30 anti-HTLV-III antibody is labelled with anti-(human IgG) antibody.

19. A method of detecting antibody against HTLV-III in a biological fluid, which comprises the steps of:

(a) incubating an immunoabsorbent according to any
35 of claims 14 to 16 with a sample of the biological fluid,

under conditions which allow anti-HTLV-III antibody in the sample to bind to the immunoadsorbent;

(b) separating the immunoadsorbent from the sample; and

5 (c) determining if antibody has bound to the immunoadsorbent as an indication of anti-HTLV-III in the sample.

20. A method according to claim 19, wherein step (c) comprises incubating the immunoadsorbent with (i) a
10 labelled antibody against antigen of the species from which the biological fluid is derived, (ii) labelled HTLV-III protein selected from R10, PB1, 590 and KH1, or (iii) labelled protein A; separating the immunoadsorbent from the labelled antibody, HTLV-III protein or protein
15 A; and detecting the label associated with the immunoadsorbent.

21. A method of detecting antibody against HTLV-III in a human serum or plasma sample, which comprises the steps of:

20 (a) incubating a bead of an immunoadsorbent according to any of claims 14 to 16 with the serum or plasma sample under conditions which allow anti-HTLV-III antibody in the sample to bind to the immunoadsorbent;

(b) separating the immunoadsorbent and the sample;

25 (c) incubating the immunoadsorbent with a labelled anti-(human IgG) antibody under conditions which allow the anti-(human IgG) antibody to bind human anti-HTLV-III antibody bound to the immunoadsorbent;

(d) separating the immunoadsorbent from the unbound
30 anti-(human IgG) antibody; and

(e) evaluating the label associated with the immunoadsorbent as an indication of the presence of antibody against HTLV-III in the sample.

22. A method according to claim 21, wherein the
35 anti-(human IgG) antibody is an animal antibody and the

serum or plasma sample is diluted with normal serum of an animal of the same species.

23. A method according to claim 22, wherein the anti-(human IgG) antibody is a goat antibody and the
5 serum or plasma sample is diluted with normal goat serum.

24. A method according to any of claims 21 to 23, wherein the anti-(human IgG) antibody is labelled with a radioisotope, an enzyme or a fluorescent compound.

25. A vaccine composition which comprises an HTLV-III
10 protein having the antigenic properties of R10, PB1, 590 or KH1, in a pharmacologically-acceptable vehicle.

26. A recombinant HTLV-III envelope protein fragment selected from R10, PB1, 590 and KH1, for therapeutic use.

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Figure 1A

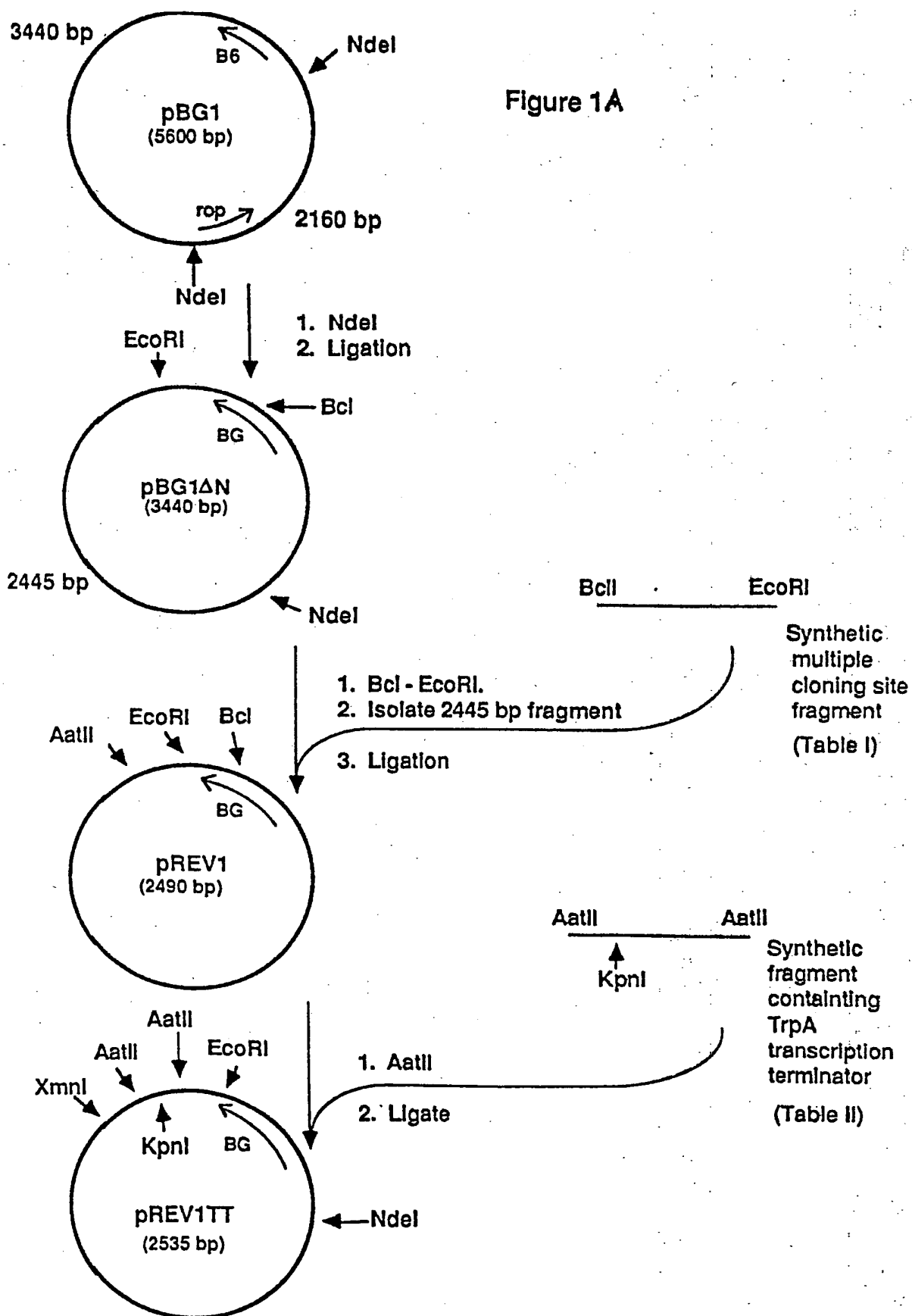
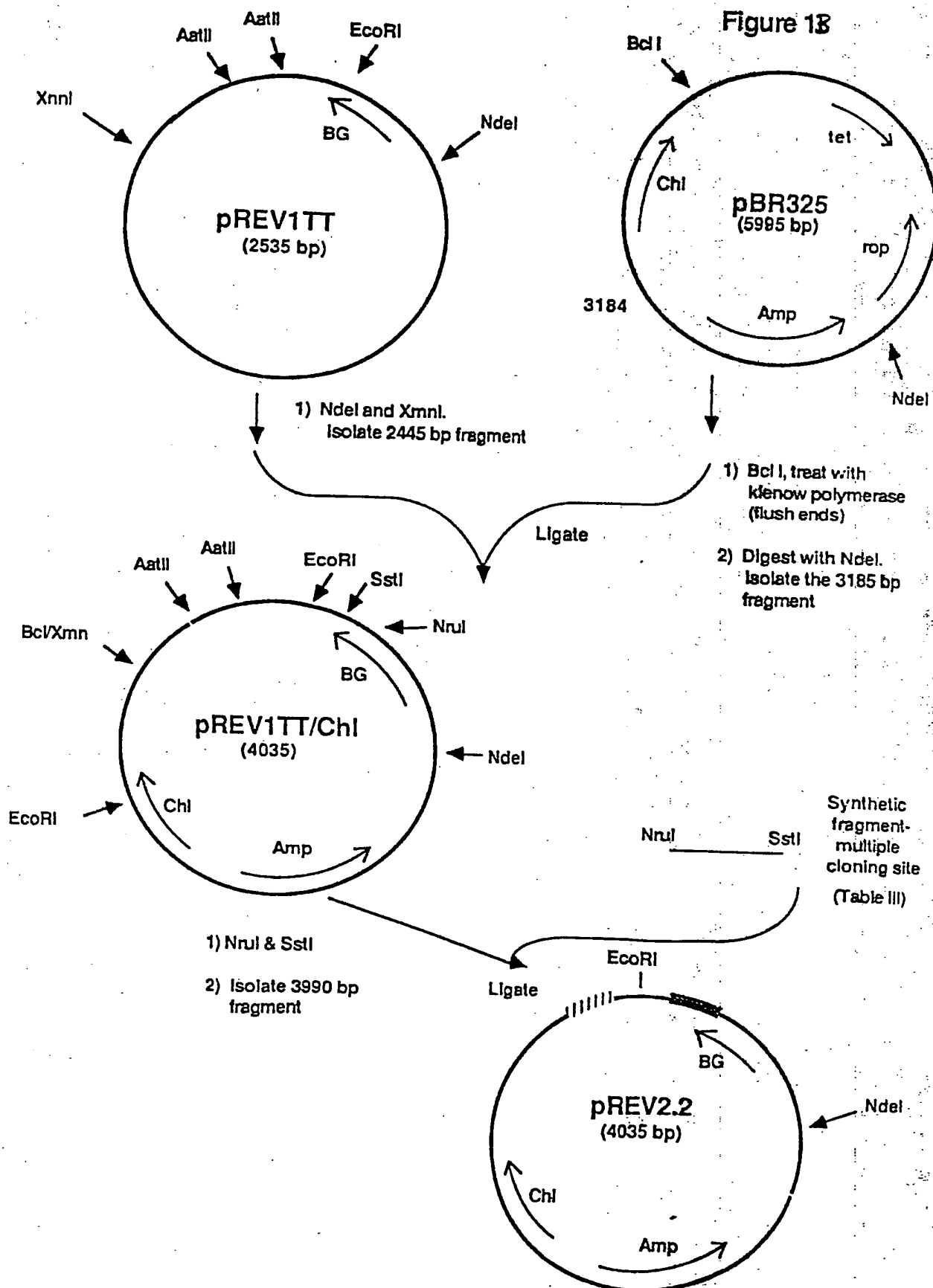


Figure 13



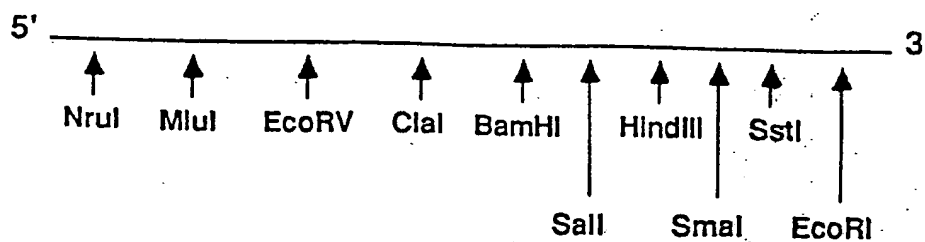
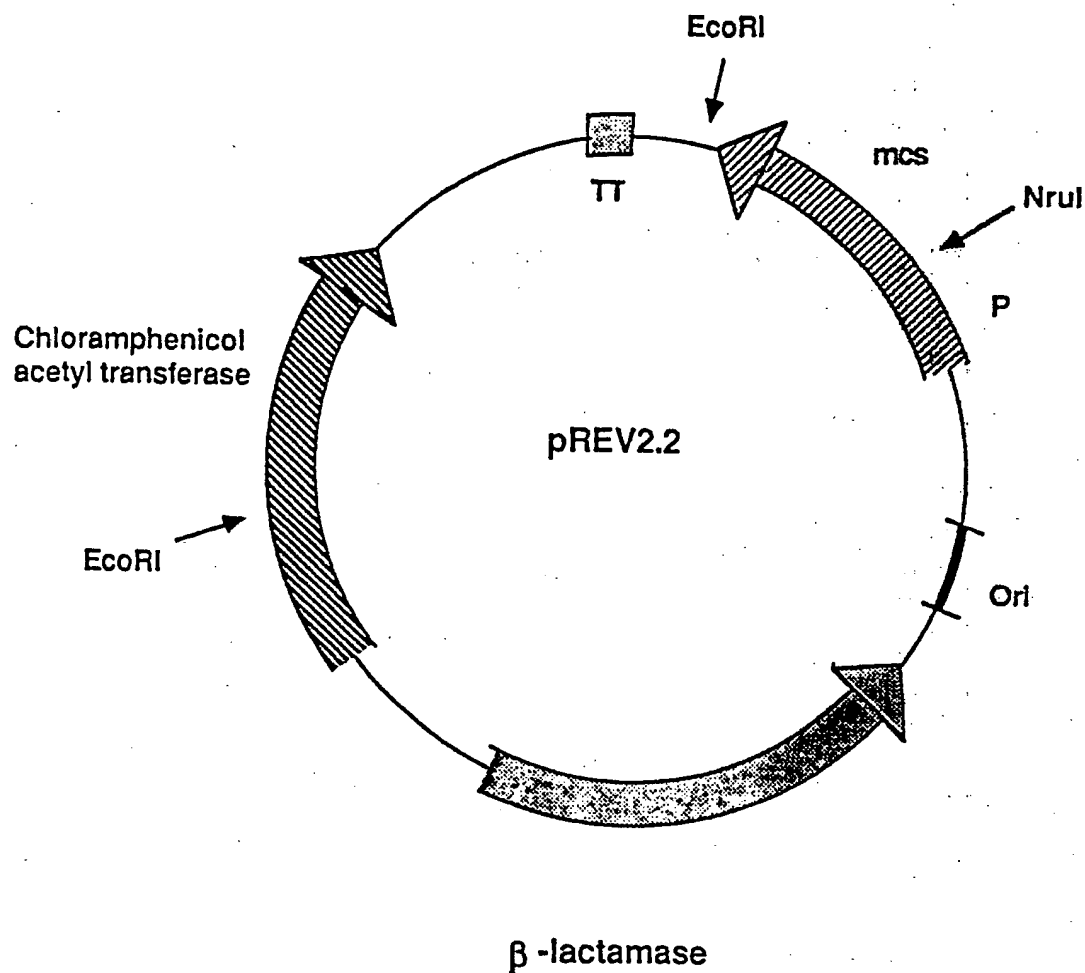
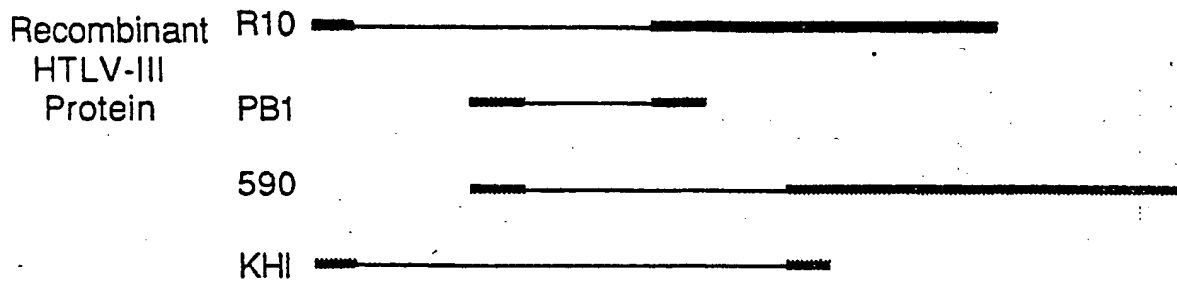
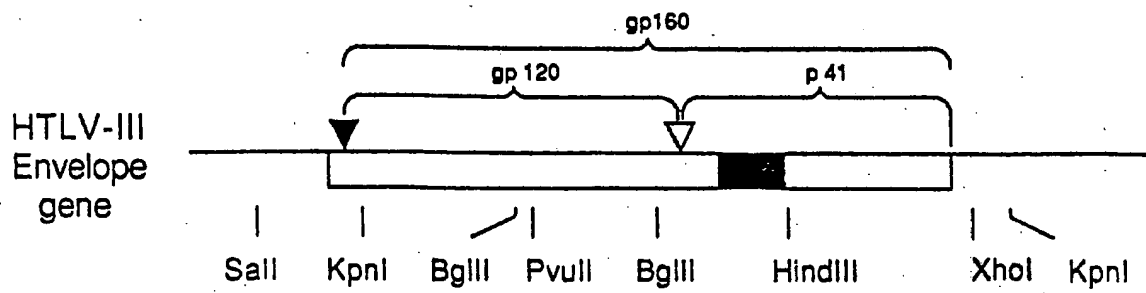


Figure 2



———— = E. coli protein sequence

Figure 3



**DECLARATION PURSUANT TO RULE 28, PARAGRAPH 4,
OF THE EUROPEAN PATENT CONVENTION**

The applicant has informed the European Patent Office that, until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, the availability of the micro-organism(s) identified below, referred to in paragraph 3 of Rule 28 of the European Patent Convention, shall be effected only by the issue of a sample to an expert.

IDENTIFICATION OF THE MICRO-ORGANISMS

Accession numbers of the deposits:

NRRL B - 18091

NRRL B - 18093

NRRL B - 18092

NRRL B - 18094

NRRL B - 18095





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Recombinant polypeptides and their uses, including assay for AIDS virus.

Novel recombinant HTLV-III envelope proteins denoted R10, PB1, 590, and KH1, are useful in the diagnosis, prophylaxis or therapy of AIDS. Protein R10 is a 95 kD fusion protein; protein PB1 is a 26 kD fusion protein; protein 590 is an 86 kD fusion protein; and protein KH1 is a 70 kD fusion protein. These proteins are considered to be especially useful to prepare vaccines for the HTLV-III virus.

EP 0 255 190 A3



European Patent
Office

CLAIMS INCURRING FEES

The present European patent application comprised at the time of filing more than ten claims.

- ☐ All claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for all claims.
- ☐ Only part of the claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims and for those claims for which claims fees have been paid.
- namely claims:
- ☐ No claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims.

X LACK OF UNITY OF INVENTION

The Search Division considers that the present European patent application does not comply with the requirement of unity of invention and relates to several inventions or groups of inventions.

namely:

See sheet -B-

- ☐ All further search fees have been paid within the fixed time limit. The present European search report has been drawn up for all claims.
- ☐ Only part of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the inventions in respect of which search fees have been paid.
- namely claims:
- ☒ None of the further search fees has been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the invention first mentioned in the claims.

namely claims: 1, 7 and partially 5-6, 11-25

DOCUMENTS CONSIDERED TO BE RELEVANT					
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 4)		
P,A	EP-A-0 199 301 (HOFFMANN LA ROCHE & CO.) * The whole document *	1, 5-7, 11-25			
	--				
P,A	EP-A-0 212 532 (SYNTEX INC.) * The whole document *	1, 5-7, 11-25			

			TECHNICAL FIELDS SEARCHED (Int. Cl. 4)		
X06X066K16NDXXK085X640C00KX07KX1R50P					
Place of search		Date of completion of the search	Examiner		
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LACK OF UNITY OF INVENTION

The Search Division considers that the present European patent application does not comply with the requirement of unity of invention and relates to several inventions or groups of inventions,

namely:

1. Claims 1,7 and partially 5-6,11-25 in so far as they relate to the below-mentioned recombinant:

DNA recombinant vector encoding for a fragment of the env gene between restriction sites Kpn I and Bg III, translated protein and uses thereof.

2. Claims 2,8 and partially 5-6,11-25 in so far as they relate to the below-mentioned recombinant.

DNA recombinant vector encoding for a fragment of the env gene between restriction sites Pvu II and Bg III, translated protein and uses thereof.

3. Claims 3,9 and partially 5-6,11-25 in so far as they relate to the below-mentioned recombinant.

DNA recombinant vector encoding for a fragment of the env gene between restriction sites Pvu II and Hind III, translated protein and uses thereof.

4. Claims 4,10 and partially 5-6,11-25 in so far as they relate to the below-mentioned recombinant.

DNA recombinant vector encoding for a fragment of the env gene between restriction sites Kpn I and Hind III, translated protein and uses thereof.

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